

Nitric Oxide in Cell Survival: A Janus Molecule

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Abstract

Nitric oxide (NO), plays multiple roles in the nervous system. In addition to regulating proliferation, survival and differentiation of neurons, NO is involved in synaptic activity, neural plasticity, and memory function. Nitric oxide promotes survival and differentiation of neural cells and exerts long-lasting effects through regulation of transcription factors and modulation of gene expression. Signaling by reactive nitrogen species is carried out mainly by targeted modifications of critical cysteine residues in proteins, including S-nitrosylation and S-oxidation, as well as by lipid nitration. NO and other reactive nitrogen species are also involved in neuroinflammation and neurodegeneration, such as in Alzheimer disease, amyotrophic lateral sclerosis, Parkinson disease, multiple sclerosis, Friedreich ataxia, and Huntington disease. Susceptibility to NO and peroxynitrite exposure may depend on factors such as the intracellular reduced glutathione and cellular stress resistance signaling pathways. Thus, neurons, in contrast to astrocytes, appear particularly vulnerable to the effects of nitrosative stress. This article reviews the current understanding of the cytotoxic *versus* cytoprotective effects of NO in the central nervous system, highlighting the Janus-faced properties of this small molecule. The significance of NO in redox signaling and modulation of the adaptive cellular stress responses and its exciting future perspectives also are discussed. *Antioxid. Redox Signal.* 11, 2717–2739.

Introduction

IN THE 1980s, nitric oxide (NO) was identified as an integral component of both the cardiovascular system and the immune response to pathogens (84). These seminal discoveries prompted an explosion in the study of nitric oxide chemistry in biologic systems, leading to one of the largest and fastest growing areas in biomedical science (131, 174). Traditional views of nitric oxide biology have been based on four key principles: (a) NO synthases [endothelial NO synthase (eNOS), neuronal NO synthase (nNOS), and inducible NO synthase (iNOS)] have unique and independent physiologic functions; (b) NO diffuses freely in biologic systems to carry out the tasks of each isoform; (c) the physiologic actions of NO are mediated primarily by cyclic guanosine monophosphate (cGMP), through the binding of NO to the heme group in soluble guanylate cyclase (sGC); and (d) the cytotoxic properties of NO are the consequence of noxious and non-specific chemistry (72). This complexity has challenged NO research related to normal physiology and disease processes. Nitric oxide, first identified as endothelium-derived relaxing factor in blood vessels (75), has been recognized, during

subsequent years, to play a pivotal role in intercellular communication, as well as in intracellular signaling, in many tissues (136).

The main targets of NO under physiologic conditions are metal centered, particularly iron in iron-heme proteins. The most striking example is the formation of a nitrosyl complex with iron (II) of sGC, inducing a conformational change of the enzyme and its activation to produce cGMP and the subsequent activation of protein kinase G (PKG) and protein phosphorylation (32). Another major pathway by which NO mediates its biologic effects (cGMP-independent pathway) is the S-nitrosylation of cysteine residues in multiple cell types and tissues (72). Alternatively, NO has been shown to mediate posttranslational modification of proteins, such as ADP ribosylation (68) and nitration of tyrosine (Tyr) residues (20). These modifications may prevent normal interactions between proteins involved in presynaptic membrane-specific interactions occurring during exocytosis or in inhibition of cell death by apoptosis (171).

A role for NO as an intercellular diffusible messenger in the brain was demonstrated for the first time a few decades ago (78). Since then, both its physiologic role, mainly related to

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regulation of neuronal proliferation/survival/differentiation as well as to mediation of synaptic activity/neural plasticity, and its contribution to several neuropathologic states have been addressed in thousands of articles (for comprehensive reviews, see 31, 57). Of particular interest is the finding that NO, known to exert an antiproliferative action toward many cell types (33), also promotes a similar effect toward neural-derived tumor cells, as well as toward neuronal precursors (156). NO itself is a poorly reactive species; however, it is able to undergo secondary reactions to form highly oxidizing and nitrating species (*viz.*, NO_2 , N_2O_3 , and ONOO^-). These secondary reactive nitrogen species (RNS) are capable of modifying a diversity of biomolecular structures in the cell. Nitric oxide is produced from L-arginine by the NO synthase (NOS) family (Figs. 1 and 4), and NO is involved in several cellular functions, such as neurotransmission, regulation of vessel tone, and immune response (32). In the central nervous system (CNS), NO production has been associated with cognitive function, because of its role in the induction and maintenance of synaptic plasticity, control of sleep, appetite, thermoregulation, and neurosecretion (Fig. 2). With regard to the peripheral nervous system, NO regulates the nonadrenergic noncholinergic relaxation of the smooth muscle in the corpora cavernosa, thus promoting penile erection, and in the gastrointestinal tract (Fig. 2). Intracellular signal-transduction pathways are involved in NO effects, with stimulation of sGC the first to be described. After this observation, many studies demonstrated that NO is able to interact with other intracellular targets, thus triggering a wide array of stimulatory or inhibitory signals (Figs. 3 and 4).

In contrast, excess NO, which can be produced by activated microglia in an uncontrolled inflammatory response (16) or by the activation of nitric oxide synthase in excitotoxicity (59), has been reported to participate in acute and chronic neuro-

degenerative diseases including stroke, multiple sclerosis, Parkinson disease (PD), and Alzheimer disease (AD) (22, 23, 194) (Fig. 4). RNS-induced modifications to proteins affect individual amino acids (primarily cysteine, tyrosine, and tryptophan), as well as the peptide backbone and metal co-factors of enzymes (Figs. 1 and 3). Recently, the term "nitrosative stress" has been used to indicate the cellular damage elicited by NO and its sequelae, mainly peroxynitrite and N_2O_3 (170). In recent years, S-nitrosylation of proteins by NO, as a cGMP-independent pathway, has been widely investigated. S-Nitrosylation is likely a prototypic redox-based signaling mechanism (99) because the S-nitrosothiols (RSNO) not only can be reduced to form thiols (Fig. 3), but can also be oxidized to form either S-glutathionylation (–SSG), cysteine sulfenic acid (–SOH), cysteine sulfinic acid (– SO_2H), or cysteine sulfonic acid (– SO_3H). The substantially different roles of these different types of modification are implicated in various physiologic and pathologic processes (88, 193). Numerous S-nitrosylated proteins have been identified *in vivo*, including serum albumin (193), hemoglobin β -subunits (81), the ryanodine-sensitive calcium-release channels (210), N-methyl-D-aspartate (NMDA) receptor (44), methionine adenosyl transferase (163), caspase-3 (126), and matrix metalloproteinases (83). We review the current understanding of the effects of NO in mammalian CNS, highlighting the Janus-faced properties of this important small molecule in neuroprotection and neurotoxicity and its significance in redox signaling and modulation of gene expression.

The NO System

Chemistry of nitric oxide

Nitric oxide (NO) is produced from the catalytic conversion of arginine to citrulline by nitric oxide synthases (NOSs) (32)

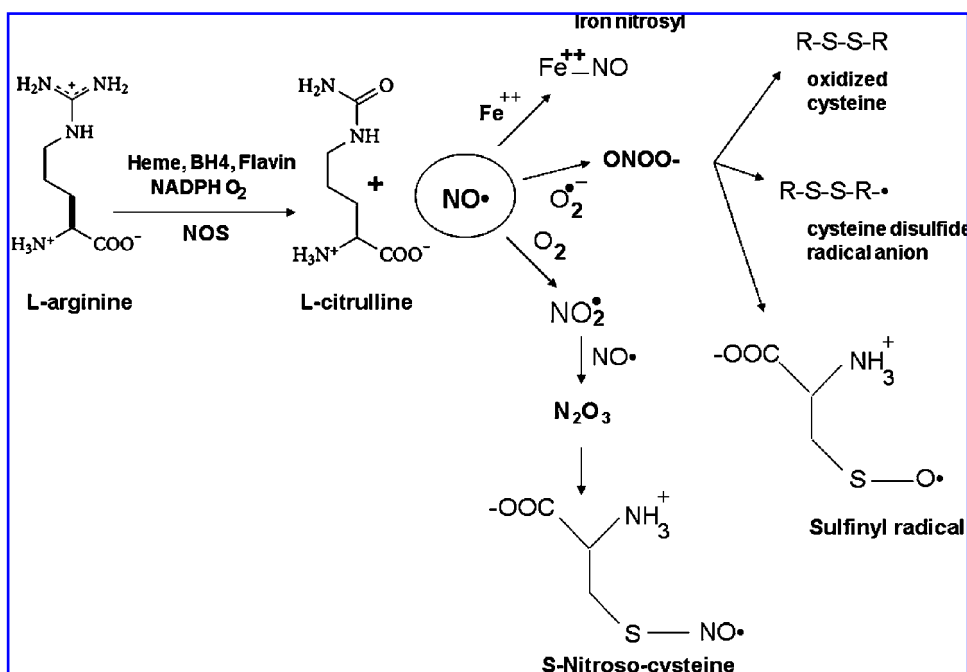


FIG. 1. The metabolic pathway leading to nitric oxide (NO) formation and consequent NO-related chemistry. Nitric oxide (NO^*) is poorly reactive with most biologic molecules, with the exception of other radical species. In the presence of oxygen, NADPH, and cofactors such as flavin mononucleotide (FMN), flavin adenine dinucleotide (FAD), heme and tetrahydrobiopterin (BH_4), nitric oxide synthase (NOS) catalyzes the oxidation of the terminal guanidinylnitrogen of the amino acid L-arginine to form L-citrulline and NO^* . At low levels, NO reacts mainly with ferrous iron in heme proteins. Higher levels may react with superoxide or oxygen, producing ONOO^- and N_2O_3 , respectively. Cysteine undergoes S-nitrosylation and oxidation

on reaction with N_2O_3 and ONOO^- , respectively. Oxidation of cysteine by ONOO^- yields disulfide bonds (with neighboring cysteines) by a mechanism with a sulfenic acid (RSOH) intermediate. In addition, homolysis of ONOO^- in either the presence or absence of CO_2 can result in a variety of products, including disulfide radical anion ($\text{RSSR}^{\cdot-}$) and sulfenyl radical (RSO^*).

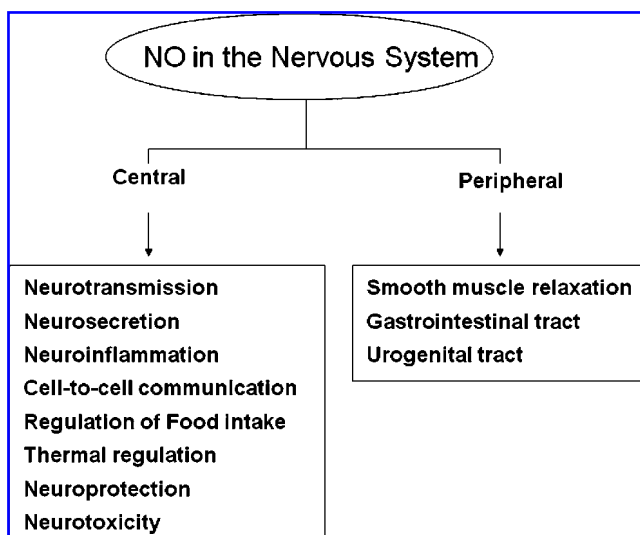


FIG. 2. Some of the main effects of nitric oxide (NO) in the central and peripheral nervous system.

(Figs. 1 and 4). Nitric oxide is neither a strong oxidant nor a strong reductant. Typically, nitric oxide does not react rapidly, with the exception of reaction with superoxide radical anion ($O_2^{\cdot-}$) (23). Nitric oxide reacts with superoxide radical anion at a diffusion-controlled rate (common among radical-radical recombination reactions) to produce peroxynitrite ($ONOO^-$) (Fig. 5A). About 15% of superoxide produced by mitochondria goes toward the formation of peroxynitrite (the other 85% forms hydrogen peroxide) (108). Importantly, $ONOO^-$ formation in mitochondria induces cytochrome *c* release, an indicator of mitochondrial distress and potential inducer of cell death, and also irreversibly blocks the respiratory chain by competing with molecular oxygen (137). Two other reactions that involve NO modification of proteins, nitrosylation and nitrotyrosylation, are important to both the physiologic and pathophysiologic roles of NO. Nitrosylation is the reaction of NO with the amino acid cysteine to form nitrosothiols on interacting proteins. Nitrotyrosylation is the reaction of the amino acid tyrosine in target proteins, with

$ONOO^-$ -derived $\cdot NO_2$ to form 3-nitrotyrosine, which has been used as a marker in neurodegeneration models (see later). Several factors influence the steady-state concentration of NO within the biologic system, including its rate of production by the various forms of NO synthase, and its rate of removal by oxidation, diffusion, NO dioxygenase, and oxy-hemoglobin (60), whereas the concentration of superoxide anion is regulated by the activity of superoxide dismutase (SOD).

Being first order with respect to both reactants, the kinetics of the reaction of nitric oxide with superoxide anion depend on the activities of NOS and SOD. Cells can regulate the concentration of superoxide anion and, hence, the intracellular formation of peroxynitrite, through cytosolic CuZnSOD and mitochondrial MnSOD. The formation of extracellular peroxynitrite is controlled by secreted or extracellular SOD (EC-SOD) (23, 24), located on the cell-surface membrane:



Peroxynitrite can exist in two forms: the nucleophilic peroxynitrite anion ($ONOO^-$), or the protonated peroxynitrous acid ($HONOO$) (2):

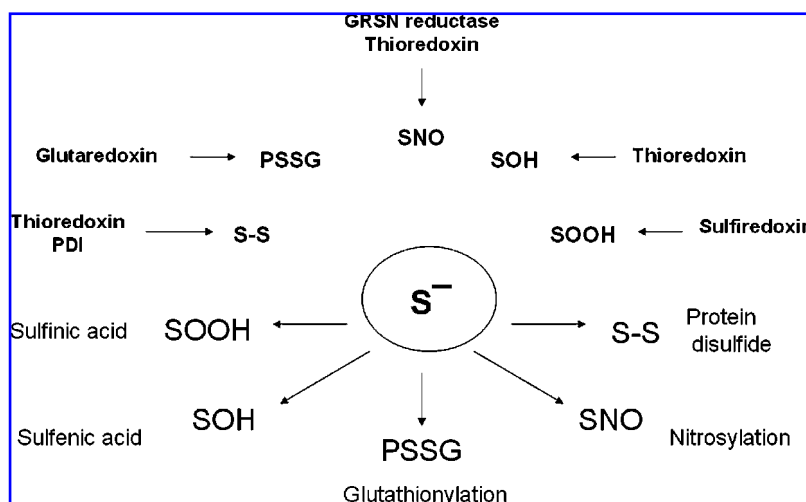


Peroxynitrite is highly reactive, with a half-life of <1 s. Peroxynitrite can undergo a variety of chemical reactions depending on its cellular environment and the availability of reactive targets, such as DNA, proteins, and lipids (23, 145). In addition to the first-order decay to form nitrate (112), peroxynitrous acid can undergo homolytic decomposition to form a hydroxyl radical and a nitrite radical (3, 10):



Peroxynitrite can react with carbon dioxide faster than the uncatalyzed decomposition of peroxynitrite; hence, the possibility exists that the *in vivo* oxidative damage attributed to peroxynitrite may actually be mediated by the reactive intermediates from the reaction of peroxynitrite and carbon dioxide, rather than by the peroxynitrite itself (62). However,

FIG. 3. Reversible modifications of reactive cysteines (S^-) and enzymes controlling reversible cysteine oxidations: S-S, disulfide; PSSG, glutathionylation; SNO, nitrosylation; SOH, sulfenic acid; SO_2H , sulfinic acid.



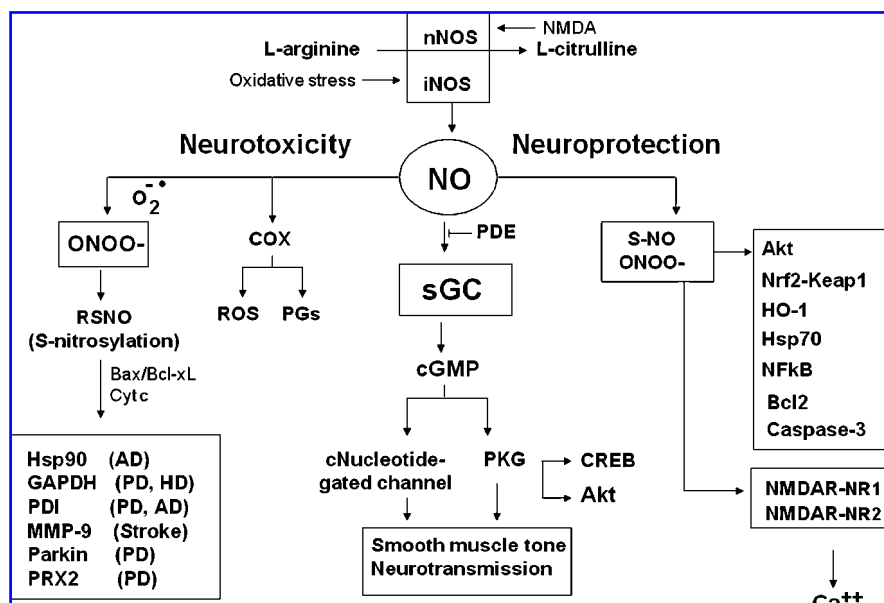
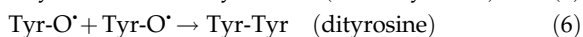
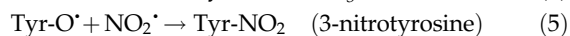
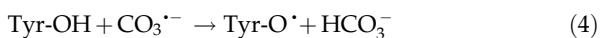


FIG. 4. The neuroprotective versus neurotoxic effect of nitric oxide. NO exerts neuroprotection at least through three mechanisms: the S-nitrosylation of the NR1 and NR2 subunits of NMDA receptor, as well as of caspase 3; the stimulation of the sGC/cGMP/PKG system; and the induction of the protooncogene Akt, as well as heme oxygenase and the Keap1/Nrf2/ARE pathway. As a consequence of the S-nitrosylating reactions, the inhibition of calcium influx through the NMDA receptor and the inhibition of caspase-3 activity occur, both leading to a decrease in cell death. The increase in cGMP levels and the consequent stimulation of PKG, as well as the induction of Akt, share the ability to activate CREB, resulting in cytoprotection. NO-mediated transcriptional activation of the Keap1/Nrf2/ARE pathway, by upregulating vitagenes such

as HO-1 and Hsp70, triggers prosurvival mechanisms. If produced in excess or in a prooxidant status of the cell, NO, by interaction with superoxide and the consequently formed peroxynitrite, produces cytotoxic effects. Consequent to the interaction between peroxynitrite and cellular components, protein nitration takes place with destruction of the cellular establishment. The S-nitrosylation of selected substrates, such as MMP-9, parkin, GAPDH, and PDI, has been hypothesized as a critical mechanism through which NO becomes neurotoxic. NO has been shown to interact with other members of the hemoprotein family, such as cyclooxygenase (COX), thus reinforcing its neurotoxic potential.

in certain instances, the presence of carbon dioxide actually inhibits the reaction of peroxynitrite with a biotarget, as is the case with the reaction of peroxynitrite with glutathione (23).

The reaction of peroxynitrite and carbon dioxide proceeds with the $ONOO^-/CO_2$ reactive pair (Fig. 5A) (122). The first intermediate in the reaction of peroxynitrite and carbon dioxide is nitrosoperoxycarbonate, which rearranges to form the second intermediate nitrocarbonate. Homolysis of nitrocarbonate is not favored but can occur to generate a nitrite radical (NO_2^{\cdot}) and a carbonate radical anion ($CO_3^{\cdot -}$) (4) (189). These nitrating and oxidizing species can then react with biomolecules, forming modifications such as 3-nitrotyrosine (5) (Fig. 5B). 3-Nitrotyrosine is a covalent protein modification that has been used as a marker of nitrosative stress under a variety of cellular conditions, especially diseased versus control states (94). Dityrosine and tyrosine peroxide may also be formed from the reactions (6) and (7) (205).

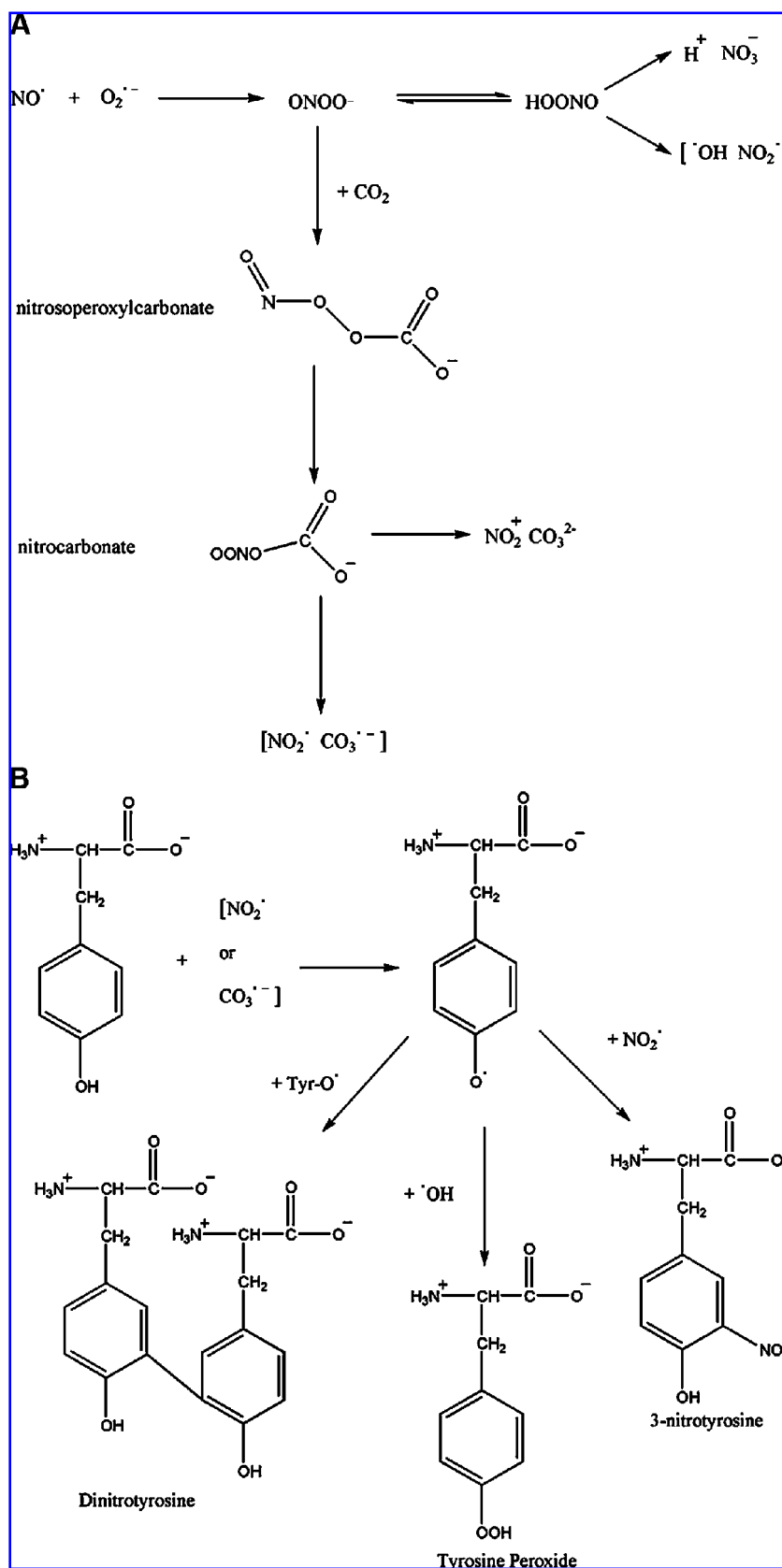


Alternatively, nitrocarbonate can undergo heterolytic cleavage, resulting in the formation of NO_2^+ and CO_3^{2-} . NO_2^+ can nitrate tyrosine residues in proteins in hydrophobic environments by electrophilic substitution (205). Figure 5A and B provide a summary of the reactions discussed earlier. Peroxynitrite can also react with sulfhydryl compounds intracellularly because of the high concentration of free thiols within the cell (99).

Reversible modifications of reactive cysteine

Proteins. Within proteins, amino acids that are targets for reversible oxidation include cysteines with a low- pK_a sulfhydryl group (98), which causes unique susceptibility to oxidation compared with the typical pK_a of nonreactive cysteines (170). Cysteine has been shown to undergo S-nitrosylation and oxidation on reaction with N_2O_3 and $ONOO^-$, respectively (Fig. 3). The individual cysteines targeted for modification may not be located within a conserved primary amino acid sequence; however, the proximal positioning of acidic and basic residues has been proposed to accelerate S-nitrosylation. Cysteines within hydrophobic regions of proteins can be targeted for nitrosylation. Cysteine can be directly oxidized by $ONOO^-$. Of all the amino acids, cysteine reacts the fastest with $ONOO^-$, with a second-order rate constant of $\sim 3 \times 10^3 \text{ M/s}$ at pH 7.4. Oxidation of cysteine by $ONOO^-$ yields disulfide bonds (with neighboring cysteines) through a mechanism with a sulfinic acid (RSOH) intermediate. In addition, homolysis of $ONOO^-$ in either the presence or absence of CO_2 can result in a variety of products, including disulfide radical anion ($RSSR^{\cdot -}$), sulfinyl radical (RSO^{\cdot}), RSNO, and R-S- NO_2 . Numerous classes of proteins contain free cysteine residues that are highly conserved across species, suggesting regulatory possibilities beyond structural roles and metal ion coordination (159). Cysteines, however, can be modified through alternative redox-based modifications (SNO, SOH, SSG, S-S), serving as molecular switches to integrate redox-based signals into specific functional responses. Recent studies have also shown that the sulfinic acid form of 2-Cys peroxiredoxins can be reduced back to the sulfhydryl in an ATP-dependent enzyme (sulfiredoxin; Srx)-catalyzed reaction (172), and a reversible thioesterification has been reported to modulate

FIG. 5. Chemistry of protein nitration. Nitric oxide is a free radical (A) that in subsequent chemistry leads to an NO_2 group on the 3-position of tyrosine. Whether using valence-bond theory (in which the NO molecule is viewed as having 11 valence electrons: 5 from N and 6 from O) or molecular orbital theory (in which an unpaired electron occupies a π^* antibonding orbital), NO has one unpaired electron. Consequently, this molecule is a free radical. Although this molecule is relatively nonreactive, in the presence of superoxide radical anion, radical-radical recombination is rapid, forming peroxynitrite. This molecule, in turn, in the presence of carbon dioxide, can lead to introduction of a NO_2 moiety on the 3-position of tyrosine (B). The *ortho*- and *para*-directing nature of the OH group in tyrosine directs the NO_2 moiety to the 3-position, because the *para*-position is already occupied in this amino acid.



transcriptional activity (86). Cysteines, in addition, are uniquely targeted by alkylating species (such as cyclopentenone prostaglandins, acrolein), by hydroxynonenal, and by acylation (99).

Like NO and SNO, H_2O_2 also oxidizes reactive cysteines, leading to the formation of SOH, which can be further oxidized to sulfinic (SO_2H) and sulfonic (SO_3H) acids (185). Sulfinic (SOH) acids also are byproducts of S-nitrosylation and targets of S-glutathionylation. In particular, S-glutathionylation, the formation of a disulfide between the cysteine of glutathione and a cysteine moiety of a protein (protein mixed disulfide, or PSSG), is believed to be a protective mechanism that prevents further oxidation to sulfinic and sulfonic acids, which are not readily regenerated (133). PSSG formation is also a consequence of denitrosylation by GSNO reductase. Recently, in HEK cells transiently expressing iNOS, it has been shown that Trx forms mixed disulfides with multiple proteins in an NO-dependent manner (11), which demonstrates the capability of Trx1 to interact with SNO-caspase-3 *in vivo* and to denitrosylate, by a mixed-disulfide intermediate mechanism, also a broad spectrum of proteins. These findings indicate that SNO-protein denitrosylation (and thus regulation of NO-based signaling) represents an additional function of the Trx system (11). Accordingly, bilirubin (BR), an endogenous molecule with strong antioxidant properties (33, 34) was shown to act as a denitrosylating agent within a concentration range of 0.1–20 μM , which corresponds to those existing within the cell (2.5 μM) or in the plasma (20 μM), increasing the release of NO from RSNO. Mass spectrometry analysis has revealed that NO binds to BR, forming an N-nitroso derivative (BR-NO). Formation of BR-NO was confirmed also in rat fibroblasts exposed to prooxidant stimuli. This finding provides novel insights into the antioxidant potential of BR, suggesting the potential usefulness as a novel biomarker of nitrosative stress (9).

Glutathione. Reduced glutathione (GSH) is a likely target of reaction with peroxynitrite, resulting in the oxidation of GSH to GSSG, which can be recycled by glutathione reductase (99). Whereas the precise physiologic mechanism of nitrosylation is unclear, it has been hypothesized that NO reacts with oxygen to form N_2O_3 , which reacts with glutathione to form the S-nitrosoglutathione (GSNO) group, which is an important intermediate in S-nitrosylation signal transduction. In 1997, Gow *et al.* (81) proposed a mechanism for the formation of S-nitrosothiols *in vivo*. The proposed mechanism involves the direct reaction of NO with free sulfhydryls to form a reactive intermediate that is reduced by an electron acceptor, such as molecular oxygen (O_2), to produce RSNO (81). It was determined that cysteine accelerates the consumption of NO under physiologic conditions. Likewise, the consumption of O_2 was determined to increase in the presence of cysteine and NO. The reaction of NO and cysteine, in the presence of CuZnSOD, produced H_2O_2 . Taken together, these results suggest that the reaction of cysteine and NO leads to the reduction of O_2 to $O_2^{\cdot-}$ and the production of a nitrosothiol. Such nitrosothiols have been shown to be involved in intracellular signaling, ion channel regulation, immunologic responses, and apoptosis (23, 24).

S-Nitrosylation. S-Nitrosylation is a ubiquitous redox-related modification of cysteine thiol by nitric oxide (NO),

which transduces NO bioactivity. Accumulating evidence suggests that the products of S-nitrosylation, S-nitrosothiols (SNOs), play key roles in human health and disease (72). S-Nitrosylation of specific cysteine residues has been detected in >100 proteins of all classes and, presumably, represents the major mechanism by which NO signals (92). Similar to phosphorylation by kinases, S-nitrosylation by NOSs influences protein activity, protein-protein interactions, and protein localization. Thus, S-nitrosylation has been implicated in transmitting signals downstream of all classes of receptors, including GPCR, RTK, TNF, Toll, glutaminergic, as well as transducing signals within intracellular compartments, such as in the case of modification of transcription factors in mitochondria and the nucleus (99). S-Nitrosylation exhibits a number of essential features that are prototypical of biologic signals: (a) temporal regulation on physiologic time scales (S-nitrosylation is stimulus coupled and rapid), (b) the existence of motifs within proteins that facilitate S-nitrosylation or provide specificity, or both, (c) colocalization of target proteins with a source of NO (*e.g.*, NOS enzyme) (97), (d) reversibility, and (e) enzymatic control [*e.g.*, GSNO reductase, thioredoxin 2 (12) controls S-nitrosothiol tone (99)]. In addition, dysregulation of protein S-nitrosylation is associated with a growing number of diseases, including asthma, heart disease, and neurodegenerative diseases (12, 146). This process, which may be mediated by NO, SNO, or several oxidants, may involve nitrosative chemistry (nitrosation refers to chemical addition of NO^+), but is not restricted to reactions of NO^+ . The term nitrosyl describes the attachment of the NO moiety to a nucleophilic group or transition metal. S-Nitrosylation is therefore the preferred terminology in biologic systems in which “-ylation” (70) is standard nomenclature for posttranslational modifications of proteins. Given these considerations, the NO-associated posttranslational modification of cysteine thiols is here referred to as S-nitrosylation.

At high concentrations, NO is oxidized by O_2 to produce N_2O_3 , a potent nitrosating agent. Other nitrosating agents include nitrosonium ion (NO^+), nitrous acidium ion ($NO-OH_2^+$), and N_2O_4 ; however, at physiologic pH, N_2O_3 is the dominant nitrosating species. N_2O_3 is formed by a second-order reaction ($k = 8.4 \times 10^6$ M/s at 37°C), which occurs in two steps: the reaction of NO with O_2 to produce NO_2 and the reaction of NO_2 with a second molecule of NO to produce N_2O_3 . NO_2 itself is a strong oxidant ($E^\circ = +1.2$ V); it does not accumulate during NO oxidation reactions in aqueous solutions. Although N_2O_3 is not a strong oxidant ($E^\circ = +0.7$ V), it is capable of nitrosating secondary amines and reduced thiol groups (170). Oxidants used in physiologic signal transduction exhibit substrate specificity and produce reversible oxidations. Highly reactive oxidant species, such as ozone, hypochlorous acid, or hydroxyl radical, oxidize biomolecules without preference or specificity, produce oxidative modifications, such as the 3-nitrotyrosine and protein carbonyls, and do not put in motion signaling events that are also not easily reduced. These protein modifications generally lead to aberrant activation of signal-transduction cascades, thus inducing pathophysiologic effects. Conversely, physiologic oxidants, such as NO/SNO, have been implicated in reversible Cys-based modifications that underlie homeostatic control of a wide array of cellular responses.

Lipid nitration. NO has a partition coefficient in octanol/water (P_{octanol}) of 5.5 (2), predisposing NO to accumulate in hydrophobic regions of the cell, such as lipid bilayers and lipoproteins. Superoxide, as it may be generated by uncoupling the electron-transport chain, particularly at complex I, or by cytosolic NAD(P)H oxidase and xanthine oxidase, can be protonated to form the neutrally charged and lipid-soluble hydroperoxyl radical. Accumulation of NO and hydroperoxyl radicals in hydrophobic environments make these areas particularly susceptible to peroxynitrite formation and subsequent damage. Once formed, peroxynitrite undergoes heterolytic cleavage, creating hydroxyl and nitrite radicals that initiate lipid peroxidation chain reactions. Hydroxyl and nitrite radicals abstract hydrogen atoms from polyunsaturated fatty acids (PUFAs) to generate carbon-centered radicals that undergo further reactions with oxygen, reactive aldehydes, neighboring lipids, and additional radical species to generate lipid hydroperoxyradicals, Michael-addition adducts, and nitrosolipids.

Emerging evidence indicates oxidized lipids as a primary target for NO nitrating effects with formation of nitro derivatives ($\text{NO}_2\text{-FA}$). These pluripotent signaling molecules are generated *in vivo* as an adaptive response to oxidative inflammatory conditions and manifest predominantly anti-inflammatory signaling reactions (7). It is also important to note that directly nitrosylated lipids have been implicated in cell-signaling processes (6). Peroxynitrite has been shown to oxidize low-density lipoproteins (LDLs) (113). Binding of nitrated LDS to macrophage-scavenging receptors leads to oxidized cholesteryl ester and foam cell accumulation, and these events are believed to occur early in atherogenesis (91). NO and its derivatives can influence eicosanoid activity and product distribution by multiple mechanisms. For example, alkyl and peroxy radical intermediates of eicosanoid synthesis react with NO at diffusion-limited rates (7).

NO also readily reacts with critical tyrosyl radical catalytic intermediates and reduced iron centers within the synthetic enzymes, thereby modulating eicosanoid formation. For instance, lipoxygenases catalyze the specific dioxygenation of PUFAs to yield reactive lipid peroxides such as leukotrienes and HETEs (7). These oxidized species are generally proinflammatory, acting as chemotactic and chemokinetic agents. Nitric oxide reversibly inhibits LOX, concomitantly being catalytically consumed through reaction with an enzyme-bound lipid peroxy radical intermediate. LOX catalytic turnover thus inhibits NO-dependent activation of sGC and acts as catalytic sink of NO. Under conditions in which NO is converted to the more powerful oxidant ONOO^- , LOX isoforms are inhibited by tyrosine nitration at concentrations as low as $1\ \mu\text{M}$ (50), although it remains to be determined whether such ONOO^- concentrations occur physiologically. Protonation of NO_2^- under acidic conditions produces nitrous acid (HNO_2 , $\text{pK}_a \sim 3$), a reaction possible in the gastric compartment (pH 1.0–5.0), phagolysosomes (pH 4.0–6.0), airways of asthmatic patients, and possibly ischemic and, hence, acidotic microenvironments.

Decomposition of HNO_2 in aqueous media proceeds by a series of reactions ultimately to yield NO_2 , N_2O_3 , and N_2O_4 , all viable candidates for mediating nitration reactions. Although acidic nitration is not kinetically rapid, the residence time of lipids, proteins, and NO_2^- in acidic tissue compartments suggests that this may be a significant pathway for

nitrating reactions. Stable, linoleic acid-derived nitrated products, with levels increasing in hyperlipidemic subjects, as well as *trans* isomers, shorter-chain nitroalkenes (putative β -oxidation products of 18 carbon and longer precursors), nitroalkenes that have been further oxidized, and nitroalkanes presumably derived from nitroalkene precursors, have been found recently in human plasma and, more generally, in healthy human specimens (7).

NO and oxidized lipids, once thought to transduce metabolic and inflammatory information *via* discrete and independent pathways, thus are recognized as interdependent regulators of the immune response and metabolic homeostasis. The relation between the two pathways extends beyond co-regulation of NO and eicosanoid formation, converging, through the nitration of unsaturated fatty acids, in the production of antiinflammatory products.

Nitric Oxide Synthase Family

Responsible for NO synthesis is the NOS family of enzymes, which, in the presence of oxygen, catalyze the conversion of L-arginine to L-citrulline and NO (Fig. 1). Recently, many tools, including NOS inhibitors, NO donors, and NO scavengers, have been discovered and developed (Table 1); their appropriate use allows researchers specifically to manipulate the NOS–NO system. Three canonic isoforms [neuronal (NOS I, or nNOS), inducible (NOS II, or iNOS), endothelial (NOS III, or eNOS)] and a significant number of spliced and posttranslationally modified variants exist. In addition, new isoforms or mitochondrial variants of NOS (mtNOS) were recently described in rat liver, thymus, and brain (69).

Neuronal NOS is expressed in neurons of both developing and mature brain of several species, including humans. Brain areas in which nNOS is highly abundant are cerebral cortex, hippocampus (in particular, the CA1 region and dentate gyrus), hypothalamus (supraoptic and paraventricular nuclei), striatum, lateral dorsal and pedunculopontine tegmental nuclei, and cerebellum (58). In the CNS, this isoform also has been found in astrocytes and cerebral blood vessels (84). In addition to this central localization, nNOS has been found in peripheral NANC neurons, which innervate the smooth muscle in the gastrointestinal tract, as well as in penile corpora cavernosa, urethra, and prostate (32, 102). Endothelial NOS (eNOS) in the brain is expressed in cerebral endothelial cells and regulates cerebral blood flow; however, a small population of neurons in the pyramidal cells of CA1, CA2, and CA3 subfields of the hippocampus and granule cells of the dentate gyrus express eNOS (32). eNOS also has been found in rat astrocytes (84).

In the periphery, eNOS has been described in vascular/sinusoidal endothelium and in the smooth muscle of rodent or human corpora cavernosa (80). Inducible NOS levels in the CNS are low, but iNOS can be induced in astrocytes or microglial cells after inflammatory conditions, viral infections, and trauma (101). In a still-evolving and somewhat controversial concept, mitochondrial NOS (mtNOS) is proposed to be bound to mitochondrial PDZ domains of COX and to complex I (73), thus favoring a steric relation between the released NO, vectorially directed to the matrix and inner membrane, and the control of respiration (38). In pathologic conditions such as extreme hypoxia, mitochondrial NO could come either from stimulated NOS or from the reduction of

TABLE 1. PHARMACOLOGIC TOOLS USED IN NO RESEARCH

Substance	Effect
L-Arginine	Substrate for NOS activity. Increases NO production
N ^G -Nitro-L-arginine methyl ester (L-NAME)	Nonselective NOS inhibitor
N ^ω -propyl-L-arginine	Highly selective and potent inhibitor of nNOS
L-Arg ^{NO2} -L-Dbu-NH2*	The most selective inhibitor of nNOS
N ⁵ -(1-iminoethyl)-L-ornithine (L-NIO)	Potent inhibitor of eNOS
N-[[3-(Aminomethyl)phenyl]methyl] ethanimidamide (1400W)	Potent, highly selective human iNOS inhibitor
1H-[1,2,4]oxadiazolo[4,3-a]quinoxalin-1-one (ODQ)	Selective inhibitor of sGC
3-(5-Hydroxymethyl-2-furyl)-1-benzyl-indazole (YC-1)	Activator of sGC
S-Nitrosothiols	Endogenous NO donors
2-Phenyl-4,4,5,5-tetramethylimidazoline-1-oxyl-3-oxide (PTIO)	A scavenger of NO ¹⁵⁷
Hemoglobin and its derivatives	An endogenous scavenger of NO ^{159,160}
Bilirubin	Endogenous scavenger of NO and RNS

NO, nitric oxide; NOS, nitric oxide synthase; RNS, reactive nitrogen species; sGC, soluble guanylyl cyclase.

nitrite by COX (37). These enzymes are bound to anchoring proteins such as caveolin in cell membrane or to heat-shock protein (Hsp) 90 or 70 chaperones (111). The interaction with these proteins may evoke different effects: (a) increased (Hsp90-NOS) or decreased (caveolin-NOS) enzyme activity, (b) modified subcellular traffic (dystrophin impedes NOS I traffic to mitochondria), and (c) participation in ubiquitination and degradation (37). Neuronal NOS and eNOS are constitutively expressed and require the formation of calcium/calmodulin complexes for their activation, whereas iNOS is the inducible form and exerts its activity in a calcium-independent manner. All three isoforms of NOS for catalytic activity need cofactors such as heme, tetrahydrobiopterin (BH4), FAD, FMN, and NADPH (31). *NOS II* gene expression is modulated by inflammatory mediators, like cytokines TNF- α , IFN- γ , and LPS, that activate transcription factors like NF- κ B or AP-1 (38). The mtNOS is nNOS- α translocated to mitochondria with specific posttranslational modifications, such as myristoylation and phosphorylation of Ser-1412 in an Akt-dependent motif (RXRXXS/T); it is constitutively expressed and Ca²⁺-dependent, and it is subjected to modulation by drugs or hormones and during development (37). The activities of cytosolic NOS isoforms are able to sustain NO cytosolic concentrations large enough to reach mitochondria. The diffusion coefficient of NO in aqueous solutions is similar to that of O₂, $\sim 4 \times 10^{-6}$ cm²/s; activation of endothelium is compatible with a concentration in the arterial wall of 2 μ M NO. Most NO binds to cytosolic compounds, like myoglobin (27). Therefore, mitochondrial NO coming from cytosolic NOS results in a considerably lower concentration, about 20–100 nM; however, it may increase by fivefold after induction of inducible NOS in endotoxemia (208). In this condition, both increased NO and NO-derived superoxide anions favor the formation of intramitochondrial peroxynitrite (37).

In mitochondria, NO reversibly binds to Cu²⁺ B center of COX and consequently inhibits electron transfer to O₂ and mitochondrial O₂ uptake. NO-dependent inhibition of O₂ use is achieved at very low physiologic NO concentrations of 50–100 nM; in these conditions, NO inhibits by half the activity of COX (165). NO inhibits COX and increases the reduction levels of the components of the electron-transfer chain, including ubiquinol and ubisemiquinone, on the substrate side of COX and reacts directly with ubiquinol to produce nitroxyl anion (NO[−]) and ubisemiquinone (165). Interestingly, the

cortex of human postmortem AD brains exhibits a loss of COX activity and increased levels of iNOS (108). Studies also have shown inhibition of mitochondrial respiratory complex I in isolated brain mitochondria and intact neurons by ONOO[−] under certain conditions. In these studies, ONOO[−] inhibited complex I only after either disruption of mitochondrial integrity or decreased reduced glutathione (GSH) levels (90). This suggests a potential role for GSH in neuroprotection in PD (108). Interestingly, presymptomatic PD patients are deficient in both GSH in the substantia nigra and complex I activity (31, 108).

NO and Neurotransmission

The first evidence of a role for NO as a neurotransmitter was reported by Garthwaite *et al.* (78), who demonstrated that stimulation of cerebellar NMDA (*N*-methyl-D-aspartate) receptors by glutamate caused the release of a diffusible molecule with strong similarities to endothelium-derived relaxation factor (EDRF) (78). Several later articles showed that NO was involved in neurotransmission in both the central and peripheral nervous systems by cGMP-dependent mechanisms (Fig. 4). Long-term potentiation (LTP) and long-term depression (LTD) of synaptic transmission are well studied models of synaptic plasticity. Several lines of evidence showed that NO, produced presynaptically or in interneurons, acts postsynaptically in cerebellar and striatal LTD, whereas the postsynaptic generation of this gaseous molecule and its presynaptic site of action characterize NO as a retrograde diffusible messenger in hippocampal and cortical LTP (13). The experimental evidence that the sGC inhibitor 1H-[1,2,4]oxadiazolo[4,3-a]quinoxalin-1-one (ODQ) inhibited, whereas the sGC activator 3-(5-hydroxymethyl-2-furyl)-1-benzyl-indazole (YC-1) enhanced NO-dependent LTP in rat hippocampal and amygdala slices, demonstrated that the NO-mediated modulation of synaptic plasticity is sGC/cGMP-dependent (15, 41).

Nitric oxide is also involved in the regulation of the release of many “classic” neurotransmitters. NO has been shown to stimulate acetylcholine release in basal forebrain and ventral striatum, not directly, but rather stimulating adjacent glutamatergic neurons (84). Basal NO concentrations have been shown to reduce, whereas high levels increase, GABA release in calcium- and sodium-dependent processes (84). Nitric oxide

donors stimulated, whereas hemoglobin, an endogenous scavenger of NO, inhibited noradrenaline and glutamate release in brain areas such as hippocampus (120). In rat striatum and medial preoptic area, NO increased both dopamine and serotonin release in a cGMP-dependent way (100, 202).

Neuroprotective versus Neurotoxic Effects of NO

NO in neuroprotection

NO confers a neuroprotective effect through multiple mechanisms. This effect has been demonstrated in several experimental models (Figs. 2 and 4). Activation of soluble guanylate cyclase by NO, through reaction with its heme center, followed by increased production of cGMP and activation of protein kinases G (PKG I and PKG II), was the first identified cellular target for transduction of NO-mediated signals (93, 144). This transduction pathway is involved in the response of many different cell types to the NO signal and can affect the function of a wide array of proteins, as well as modulate the function of other cellular messengers, such as cAMP and calcium (84). As noted earlier, NO also reacts with thiol groups of amino acid residues, especially with cysteine, forming *S*-nitrosylated derivatives (193). Both nitration and *S*-nitrosylation affect the function of the target proteins, ensuring that a multiplicity of cell-specific effects stem from the same initial signaling molecule (84). The two main modalities able to elicit cellular responses to NO (*i.e.*, through cGMP messenger or through conformational modification of protein function by direct chemical reaction) are not mutually exclusive, even if they may preferentially occur at different concentrations of NO (52).

NO and redox signaling

Signaling mechanisms adopted by regulatory proteins to control gene expression in response to increased levels of reactive oxygen and nitrogen species are common in prokaryotes. Gene activation by oxidative stress was first described in bacteria in which regulatory proteins such as OxyR and SoxR were discovered to activate the expression of antioxidant and stress-response genes (87). The activity of OxyR is turned on in response to hydrogen peroxide and *S*-nitrosothiols, and the first model postulated that OxyR is activated through the formation of a disulfide bond between two of its cysteine residues, C199 and C208, and is deactivated by enzymatic reduction of this disulfide bond with glutaredoxin 1 (219, 220). Recent studies suggested that modifications of a single cysteine (C199) in response to different stresses to (a) sulfenic acid, C199-SOH, by oxidative stress; (b) *S*-nitrosothiol, C199-SNO, by nitrosative stress; or (c) mixed disulfide with glutathione (C199-S-S-G) by disulfide stress may represent the sensor mechanism (105, 159). The expression of OxyR-dependent genes renders the bacteria more resistant to oxidant damage (167). The *Escherichia coli* SoxR and SoxS proteins (which principally mediate responses to superoxide) have also been implicated in responses to nitric oxide (190). SoxR activation occurs when its binuclear iron-sulfur [2Fe-2S] clusters undergo a reversible one-electron oxidation. Nitrosylation of the [2Fe-2S] clusters to form protein-bound dinitrosyl-iron-dithiol adducts has been demonstrated after treatment of intact bacteria or purified SoxR with nitric oxide (64). SoxR turns on the transcription of SoxS, which in turn

activates the expression of the SoxRS regulon-encoding proteins with protective functions, including sodA (manganese-containing superoxide dismutase), zwf (glucose-6-phosphate dehydrogenase), fldA and fldB (two distinct flavodoxins), and fpr (NADPH-ferredoxin reductase).

Cyclic AMP response element-binding protein and Akt

In primary rat cerebellar granule cells cultured for 7 days, inhibition of NO synthesis resulted in a significant increase in apoptotic cell death through the activation of caspase-3. Interestingly, apoptosis secondary to NO deprivation in these cells was mimicked by the sGC inhibitor ODC and was reversed by treatment of these neurons with NO donors or cGMP analogues (47). With this experimental system, the intracellular pathways through which NO exerts neuroprotective effects also were delineated (48), where the involvement of the kinase Akt and the transcription factor CREB in the survival pathway elicited by NO in cerebellar granule cells was clearly demonstrated (49); notably, both proteins are important as signal transducers of neurotrophin-mediated survival and protection against various neurodegenerative challenges, and this similarity contributes to the neuroprotective role of NO (32, 52) (Fig. 4). cAMP response element-binding protein (CREB) is a leucine-zipper transcription factor, which activates transcription of a large set of genes having the consensus sequence CRE in their promoters and whose functions in neural cells have been extensively studied during the past decade. This stimulus-induced transcription factor is activated through phosphorylation at Ser-133 by a large array of extracellular signals essential for neuronal function, such as neurotrophins and neurotransmitters, and regulates the expression of genes that in turn promote neuronal survival, protection from neurodegenerative insults, neurogenesis, synaptic plasticity, and memory formation and consolidation (138).

The contribution of NO to the mechanisms of synaptic plasticity, such as hippocampal LTP, and to the learning and memory has been demonstrated by several researchers and is supported by genetic or pharmacological manipulation of the NO/cGMP system (189). Although some of these effects are related to retrograde messages conveyed by diffusion of NO from the postsynaptic neurons, other effects have been linked to the activation of CREB-mediated transcription in the postsynaptic neuron (52). Recent evidence has clearly demonstrated that NO promotes neuron survival through regulation of CREB transcriptional activity. Pharmacologic dissection of the cellular pathway elicited by NO pointed to the essential role of the cGMP/PKG system and at the increased expression of the antiapoptotic gene *Bcl-2* in survival promotion (49). Further refinements of this finding are supported by recent studies indicating that, instead of being constitutively bound to CRE sequences, CREB binds to the DNA consensus sequences of several genes through an NO-dependent pathway targeted to *S*-nitrosylation of nuclear proteins (173).

Neuroprotection through S-nitrosylation

Similar to phosphorylation by kinases, *S*-nitrosylation by NOSs influences protein activity, protein-protein interactions, and protein localization. *S*-Nitrosylation thus serves as the fundamental redox-based signal, integrating the NO signaling by guanylate cyclase. *S*-Nitrosylation has been

implicated in transmitting receptor-mediated signals, as well as transducing, through modification of transcription factors, signals within mitochondria and the nuclear compartment (92). NO also confers neuroprotection in the NMDA-mediated neurotoxicity model, in which prolonged stimulation of the NMDA receptor causes excitotoxic cell death (32). NO protects against such excitotoxicity by S-nitrosylating the NR1 and NR2 subunits of the NMDA receptor (98, 117), which reduces the intracellular Ca^{2+} influx responsible for neuronal death (116).

Interestingly, prolonged nNOS stimulation, which occurs in response to sustained NMDA-receptor activation, generates superoxide radicals, which, in turn, react with NO to form peroxynitrite (143). Consequently, NO formed during excessive NMDA activation S-nitrosylates the NMDA subunits, thereby diminishing either the formation of peroxynitrite or Ca^{2+} influx to promote neuronal survival (Fig. 4). NO can also confer cytoprotection through the inhibition of caspase activity by S-nitrosylation of the catalytic-site cysteines (133, 118). S-Nitrosylation has been shown to reduce the activity of caspases in several cell lines, including neurons (126, 201, 203). Cortical neurons treated with several NO donors, including S-nitrosothiols, exhibited a significant reduction in staurosporin-induced caspase-3 and caspase-9 activation, probably owing to the NO-mediated S-nitrosylation of the cysteine residue in the catalytic site of these caspases. In addition, NO treatment inhibited the appearance of the classic apoptotic nuclear morphology (221). Surprisingly, caspase-3 and caspase-9 inhibition by NO was not paralleled by a significant increase in neuronal cell viability, which therefore implies the occurrence of an alternative, caspase-independent, form of cell death in neurons exposed to NO, in accordance with previous findings (133).

Neuroprotection through transcriptional control of gene expression

It has been demonstrated that in mammalian cells, NO induces temporally distinct waves of gene activation. Whereas NO may affect gene transcription by controlling the methylation of CpG islands in the promoter regions of some genes, most of its activity in regulating gene expression occurs through regulation of transcription factors (178). Nuclear factor- κ B (NF- κ B) is an important transcription factor whose biologic activities are influenced by NO. Originally found to be active in responses of immune cells, such as inflammatory responses, by inducing the expression of cytokines and adhesion molecules, this transcription factor is particularly relevant for NO-related modulation of gene expression, as ample evidence exists for its regulation through mechanisms of nitrosylation and nitration (163, 198).

In most mammalian cells, NF- κ B is constitutively expressed in the form of the p50/p65 heterodimer, and p50 S-nitrosylation at Cys-62 residue inhibits gene transcription, dependent on NF- κ B binding at the DNA promoter site (164). Nitric oxide can regulate the DNA-binding and transcriptional activity of NF- κ B either directly, by interacting with the factor itself, or indirectly through its endogenous inhibitor I κ B proteins (51, 52). In addition, the NO-dependent decreased phosphorylation and proteasomal degradation of I κ B seems to play a critical role in the cellular mechanisms leading to NF- κ B activation (129, 160). A novel mechanism for NO-mediated

inactivation of NF- κ B transcriptional activity, targeted on p65 regulation, was recently described in cell lines (52). Exposure of cells to sodium nitroprusside resulted in rapid inactivation of NF- κ B, an effect that was reverted by providing the cells with a peroxynitrite scavenger, thus suggesting the presence of a mechanism based on tyrosine nitration. Further chemical analysis through mass spectrometry revealed nitration on Tyr-66 and Tyr-152 residues of p65 (162). Centered on activation of nitrosative conformational changes of proteins, complementary cellular transduction pathways have been recently described in cortical neurons, which include brain-derived neurotrophic factor (BDNF)-induced S-nitrosylation of histone deacetylase 2 (HDAC2) and the Akt/PKB pathways (49). BDNF-induced S-nitrosylation of HDAC2 has been shown to result in histone modifications and activation of pro-survival genes, such as *Fos*, *Vgf*, and *Egr1* genes (150). Similarly, the antiapoptotic kinase Akt is activated by phosphorylation, promoted by phosphatidylinositol-dependent kinase 1 (PDK1). This kinase, in turn, is activated by the effector phosphatidylinositol-3,4,5-trisphosphate (PIP3), which is the product of PI3K activity. Activation of AKT by phosphatidylinositol 3-kinase (PI3K) serves as a multifunctional regulator of apoptotic cell death, cell growth, and glucose metabolism. With respect to neuronal cell function, AKT has been shown to be required for prevention of apoptosis and promotion of cell survival through the phosphorylation of the proapoptotic Bcl-2 family protein, Bad (56), and procaspase 9 (36). Incubation of primary cortical neurons in culture with peroxynitrite rapidly triggers phosphorylation of Akt at Ser473 and protects neurons against the apoptotic death caused by etoposide. Such effects were abolished by inhibition of PI3K activity with wortmannin, or by Akt shRNA knockdown, suggesting that the PI3K/Akt pathway mediated the antiapoptotic effect of peroxynitrite (61).

Neuroprotection through overexpression of heme oxygenase and the Keap1/Nrf2/ARE pathway

Eukaryotes are equipped with sophisticated protective mechanisms against environmental challenges. Typical examples are proteins such as heme oxygenase 1 (HO-1), thioredoxin, thioredoxin reductase, superoxide dismutase, glutathione S-transferases (GST), and NAD(P)H:quinone oxidoreductase 1 (NQO1), whose gene expression is upregulated in response to a variety of chemical and physical stresses (34, 35). The induction of heme oxygenase-1 is considered an early event in the cellular response to oxidative stress and exerts a neuroprotective function (35, 40). Moreover, under prooxidant conditions, upregulation of iNOS occurs in the brain and is followed by HO-1 induction in rat astrocytes and microglia, as well as in the hippocampus (106). The upregulation of heme oxygenase-1 protein and the following increase in biliverdin, which is further reduced by biliverdin reductase into the antioxidant and antinitrosative molecule bilirubin, can be considered a secondary mechanism through which NO can exert neuroprotective effects (21, 27).

The notion that the antioxidant protein heme oxygenase could "sense" NO and thus protect against ROS and RNS insults is supported by the following findings: (a) NO and NO-related species induce HO-1 expression and increase heme oxygenase activity in human glioblastoma cells, hepatocytes, and aortic vascular cells; (b) cells pretreated with

various NO-releasing molecules acquire increased resistance to H_2O_2 -mediated cytotoxicity at the time heme oxygenase is maximally activated; and (c) bilirubin, one of the end products of heme degradation by heme oxygenase, protects against the cytotoxic effects caused by strong oxidants such as H_2O_2 and $ONOO^-$ (141). The concept that NO and RNS can be directly involved in the modulation of HO-1 expression in eukaryotes is based on the evidence that different NO-releasing agents can markedly increase HO-1 mRNA and protein, as well as heme oxygenase activity, in a variety of tissues, including brain cells (182). In rat glial cells, treatment with lipopolysaccharide (LPS) and interferon- γ (IFN- γ) results in a rapid increase in both iNOS expression and nitrite levels followed by enhanced expression of HO-1 protein. In the same study, the presence of NOS inhibitors suppressed both nitrite accumulation and HO-1 mRNA expression (26).

Modulation of HO-1 mRNA expression by iNOS-derived NO after stimulation with LPS has also been reported in different brain regions, particularly in the hippocampus and substantia nigra in the *in vivo* rat model of septic shock (183). Moreover, the early increase in iNOS protein levels observed in endothelial cells exposed to low oxygen tension seems to precede the stimulation of HO-1 expression and activity, an effect that appears to be finely regulated by redox reactions involving glutathione (140, 215). Taken together, these findings point to the central role of the NO as a signaling molecule that, by triggering expression of cytoprotective genes such as HO-1, may lead to adaptation and resistance to subsequent, eventually more severe, nitrosative and oxidative stress insults (35).

Thus, a direct interaction of NO with selective chemical sites localized in transcription factors that can be activated through nitrosative reactions could effectively contribute to the enhancement of both HO-1 gene expression and stress tolerance. Recent knowledge concerning the modulation (by altering the thiol redox state) of the activity of several transcription factors that recognize specific binding sites within the promoter and distal enhancer regions of the *ho-1* gene include Fos/Jun [activator protein-1 (AP-1)], nuclear factor- κ B (NF- κ B), and the more recently identified Nrf proteins (8, 33). Both AP-1 and NF- κ B contain cysteine residues whose interaction with oxidative or nitrosative species might be crucial for determining the binding activity to DNA (141). Data in the literature show that NO can either activate or inhibit these transcription factors, and that in many circumstances, activation

depends on the reversibility of the posttranslational modification elicited by the various RNS (27, 175).

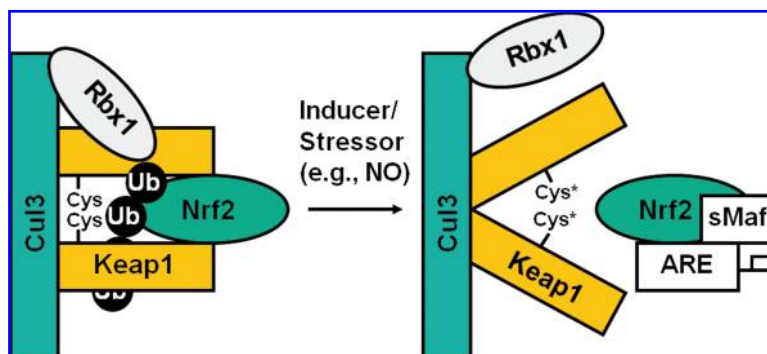
Recently, accumulating experimental evidence suggests that one of the protective functions of NO could be mediated through transcriptional activation of the Keap1/Nrf2/ARE pathway (Fig. 6), a quintessential cytoprotective pathway in eukaryotes. This pathway is composed of three major elements: (a) the sensor protein, Keap1 (Kelch-like ECH-associated protein 1) (96); (b) transcription factor Nrf2 (nuclear factor erythroid 2-related factor 2) (95, 135); and (c) specific promoter sequences ARE/EpRE (antioxidant/electrophile response element, consensus sequence 5'-A/CTGAC/GNNNGCA/G-3') (74, 148, 176, 177, 198) present in the upstream regulatory regions of a battery of >100 cytoprotective genes, among which are HO-1, thioredoxin, thioredoxin reductase, GST, and NQO1. Activation of the Keap1/Nrf2/ARE pathway leads to rapid adaptation to a variety of stress conditions and promotes cell survival (103, 110, 139).

Several models have been proposed to explain how this pathway operates under both basal and stress conditions, and they were recently comprehensively reviewed by Hayes and McMahon (89). Under basal homeostatic conditions, the homodimeric sensor protein Keap1 binds one molecule of transcription factor Nrf2 and targets it for ubiquitination and subsequent proteasomal degradation (132). Keap1 is a multidomain protein equipped with several protein-protein binding sites (66). Thus, in addition to Nrf2, Keap1 binds to Cullin 3 (Cul3), and, in doing so, Keap1 presents Nrf2 for ubiquitination to the Cullin 3 (Cul3)-Ring box 1 (Rbx1) E3 ubiquitin ligase complex (54, 76, 109, 217). A critical feature of the Keap1 protein is that it contains highly reactive cysteine residues that serve as sensors for various stressors/inducers of the Keap1/Nrf2/ARE pathway (65, 66, 207). Depending on the inducer, distinct cysteines are modified, most frequently by participating in Michael addition or oxidation-reduction reactions. We and others reported that C151, C273, and C288 are of particular importance in the sensor function of Keap1 (65, 216). Inducer sensing by Keap1 ultimately leads to stabilization of the transcription factor, allowing Nrf2 to bind (in heterodimeric combinations with small Maf proteins) to the antioxidant response elements of its target genes and to activate their transcription. This transcriptional upregulation results ultimately in enhanced cytoprotection (67, 199, 200).

In human neuroblastoma cells, nitric oxide was shown to cause nuclear accumulation of Nrf2, binding to the ARE, and

FIG. 6. The Keap1/Nrf2/ARE pathway.

Under basal conditions, transcription factor Nrf2 is bound to a dimeric partner protein Keap1. Keap1 also associates with Cullin 3 (Cul3), and, in doing so, Keap1 presents Nrf2 for ubiquitination to the Cullin 3 (Cul3)-Ring box 1 (Rbx1) E3 ubiquitin ligase complex. As a result, Nrf2 undergoes ubiquitination and proteasomal degradation. Inducers/stressors modify highly reactive (sensor) cysteine residues of Keap1, resulting in loss of its ability to target Nrf2 for ubiquitination. Consequently, Nrf2 is stabilized, binds to the ARE (in heterodimeric combinations with a small Maf transcription factor), and activates transcription of cytoprotective genes. Cys, sensor cysteine; Cys*, modified sensor cysteine. (For interpretation of the references to color in this figure legend, the reader is referred to the web version of this article at www.liebertonline.com/ars).



upregulation of Nrf2-dependent genes, and this induction counteracted NO-induced apoptosis (63). RNA interference, or transfection with a dominant negative Nrf2 construct, resulted in reduced levels of cytoprotective genes and sensitization to NO-induced apoptosis, whereas overexpression (stable transfection) of Nrf2 was protective. Treatment with NO-donating aspirin also increased the levels of several Nrf2-dependent genes in human colon (HT-29) and murine liver (Hepa1c1c7) adenocarcinoma cells, as well as *in vivo* in the liver and intestine of *Apc^{Min/+}* mice (77), and inhibited by about 60% the formation of intestinal tumors in this mouse model (209). Exposure of cardiac cells to nitric oxide donors in combination with hemin increased HO-1 mRNA and protein expression under both normoxic and hypoxic conditions (147). The HO-1 increase was accompanied by elevated nuclear levels of Nrf2, but this elevation was abolished by *N*-acetylcysteine. Similarly, exposure to nitric oxide donors increases the expression of Nrf2-dependent genes (e.g., HO-1, γ -glutamylcysteine synthetase) in vascular endothelial cells (18) and smooth muscle cells (119). Thus, in primary cultures of bovine aortic endothelial cells, nuclear levels of Nrf2 were increased after exposure to spermine NONOate, and this increase was accompanied by elevation in HO-1 expression and total glutathione levels (18). Inhibitors of ERK, p38 MAPK, as well as pretreatment with *N*-acetylcysteine, decreased the NO-dependent stabilization of Nrf2 and upregulation of HO-1, suggesting the importance of both phosphorylation and redox mechanism(s). In rat aortic smooth muscle cells, nitric oxide donors (spermine NONOate, *S*-nitroso-*N*-acetylpenicillamine, sodium nitroprusside, Sin-1, or Angeli's salt) increased the levels of Nrf2, the binding of Nrf2 to the promoter of HO-1, and the expression of HO-1 (119). Mutations in the ARE of HO-1 or overexpression of a dominant-negative Nrf2 construct abrogated the NO-dependent upregulation of HO-1. In this study, inhibitors of ERK or p38 MAPK had no effect on the NO-mediated nuclear accumulation of Nrf2. In contrast, pretreatment with ascorbate or *N*-acetylcysteine reduced the NO-mediated increase in Nrf2 levels, suggesting the involvement of redox mechanism(s).

Because Keap1 is an essential regulator of Nrf2 levels in the cell and its highly reactive cysteine residues are the sensors for inducers of Nrf2-dependent genes, Keap1 is a good candidate to serve as a sensor for nitric oxide. As discussed earlier, *S*-nitrosylated proteins can form when a cysteine thiol reacts with nitric oxide in the presence of an electron acceptor (e.g., molecular oxygen) to form an *S*-NO bond (79, 180) (Fig. 1). Furthermore, *S*-nitrosylation is now recognized as an important mechanism for the posttranslational regulation of many proteins, thus conveying the effects of nitric oxide on signal transduction (Fig. 4) (92).

Although their exact chemical nature has not been identified, reversible cysteine modifications (that could include *S*-nitrosothiols, disulfide bonds, sulfenic acid derivatives) of both overexpressed and endogenous Keap1 were reported to occur when HEK293H cells were treated with spermine NONOate or *S*-nitrosocysteine (19). Recently, the formation of a nitrated derivative of cGMP, 8-nitroguanosine 3',5'-cyclic monophosphate (8-nitro-cGMP) was reported when RAW 264.7 cells were stimulated with LPS and IFN- γ to upregulate transcriptionally iNOS and to produce nitric oxide (181). Because of the electrophilic nature of 8-nitro-cGMP, the possibility that it might react with cellular thiols to form

8-thioalkoxy-cGMP (8-RS-cGMP) adducts was explored. Under these conditions of nitric oxide production, 8-Cys-cGMP adducts in endogenous Keap1 were identified, and the reaction was termed *S*-guanylation (Fig. 7). NO-dependent *S*-guanylation of Keap1 also was found in murine peritoneal macrophages after infection with *Salmonella enterica* var. Typhimurium. *In vitro*, recombinant Keap1 was shown to be highly susceptible to *S*-guanylation induced by 8-nitro-cGMP, even in the presence of excess (>3,000-fold) GSH.

Nitric Oxide in Neurodegeneration

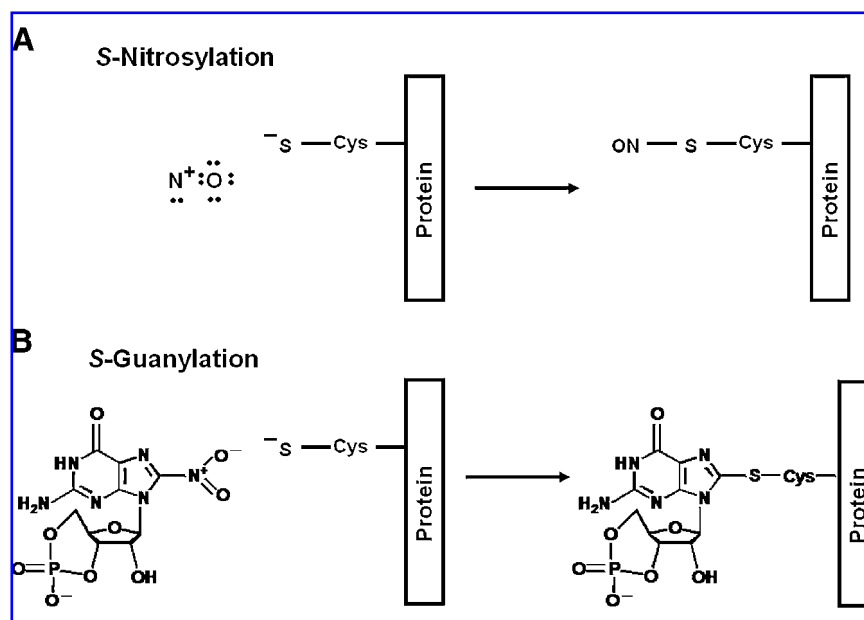
Although NO has many important and beneficial physiologic functions, it also can play a role in neurodegenerative disease pathology (31). In these diseases, NO is produced in excess by iNOS induction owing to the proinflammatory response, which is a common feature of neurodegenerative disorders (108). Moreover, NO is much more harmful under pathologic conditions that involve the production of reactive oxygen species (ROS), such as superoxide anion and the formation of peroxynitrite (34) (Fig. 4).

Two important properties of NO that may contribute to its pathologic functions are its ability to modify proteins through nitrosylation and nitrotyrosination and its ability to react with oxygen to form RNS. Accordingly, studies indicate that nitrosylation and nitrotyrosination of many different proteins play a role in the pathology of multiple neurodegenerative diseases. The formation of nitrotyrosine, as a marker of nitrosative stress, has been documented in AD and PD (27, 29, 30, 33, 130, 195, 196).

Glutamate excitotoxicity is caused by overstimulation of synaptic glutamate receptors that results in excessive Ca^{2+} influx and subsequent neuronal injury. Excitotoxicity is a common event in many neurodegenerative disorders, including AD. NO can both protect and sensitize cells to excitotoxic cell death through a mitochondrial pathway. Specifically, under normal physiologic conditions, NO *S*-nitrosylates NMDA receptors, blocks Ca^{2+} influx, and promotes cell survival. In immature neurons, however, mtNOS is responsible for the majority of NOS activity. In these conditions, Ca^{2+} enters mitochondria and stimulates NO production by mtNOS. As a consequence, inhibition of the respiratory chain occurs, which reduces the mitochondrial membrane potential, collapses the ion gradient, and decreases entry of Ca^{2+} into the mitochondria. Decreased Ca^{2+} influx causes a decrease in mtNOS activity and NO production, promoting cell survival. Conversely, in mature neurons, when cytosolic nNOS is the primary producer of NO, Ca^{2+} entry through overactive NMDA channels stimulates nNOS; thus, NO can then enter the mitochondria, directly inhibiting complex IV (cytochrome *c* oxidase) of the respiratory chain, which leads to a block of ATP production and eventual cell death due to energetic failure. In contrast to immature neurons, in mature neurons, no feedback loop is present to protect the cell from damage, and degenerative cell death ensues.

Furthermore, NO has been shown to activate both the constitutive and inducible isoforms of cyclooxygenase, which are upregulated in brain cells under proinflammatory conditions (125). During the catalytic cycle of cyclooxygenase, the release of free radicals and the formation of prostaglandins occur, two events closely related to the development of neuroinflammation (43). Interestingly, the upregulation of induc-

FIG. 7. Two possible NO-dependent modifications of cysteine residues in proteins. (A) *S*-Nitrosylation. Cysteine thiolate anion donates an electron pair to the nitrosonium cation, an electrophile that is formed from nitric oxide in the presence of molecular oxygen (180). (B) *S*-Guanylation. Cysteine thiolate anion reacts with the electrophilic 8-nitro-cGMP that is formed in cells in the presence of nitric oxide (181).



ible cyclooxygenase has been found in the brain of patients affected by AD, and it is considered a marker of the progression of dementia in AD; thus, activation of inducible cyclooxygenase can be considered an indirect way for NO to induce neurotoxicity.

Alzheimer's disease

Recently, a relation between glyceraldehyde-3-phosphate dehydrogenase (GAPDH) and the β -amyloid precursor protein (betaAPP) in relation to the pathogenesis of AD was suggested. Decreased dehydrogenase activity of cerebral glyceraldehyde-3-phosphate dehydrogenase has been described in different animal models of AD (185). In addition, recent evidence indicates that polymorphic variation within GAPDH genes is associated with an elevated risk of developing AD (55). GAPDH not only is important in ATP production, but also has a role as nitrosative stress sensor. Active-site cysteine 149 may undergo several modifications on reaction with NO (and RNS), which result in a reversible or irreversible inhibition of GAPDH enzymatic activity, depending on the severity of the prooxidant stimulus. As a consequence of this inhibition, the glucose metabolism is shifted toward the pentose phosphate shunt, which produces NADPH necessary to glutathione reductase and uncouples glucose metabolism from the production of ATP and oxidative intermediates. Glyceraldehyde 3-phosphate dehydrogenase also undergoes *S*-nitrosylation in neurons. In particular, *S*-glutathionylation after nitrosylation represents the effective inhibitory effect on GAPDH activity (197).

In addition to its well-known role in glycolysis, GAPDH contributes to nuclear signaling in apoptosis (108). *S*-Nitrosylation of GAPDH terminates its enzymatic activity and allows binding of GAPDH to Siah1, an E3 ubiquitin ligase. Siah1 has a nuclear localization signal and carries GAPDH to the nucleus. GAPDH stabilizes Siah1 in the nucleus and allows degradation of nuclear proteins through ubiquitination (108).

Protein-disulfide isomerase (PDI) is an endoplasmic reticulum (ER)-associated chaperone protein that prevents neurotoxicity caused by ER stress and protein misfolding (204) and can also function as an NO receptor or donor, depending on the cellular context (187). PDI is nitrosylated both in AD and PD. Both PD and AD patient postmortem brains exhibit increased levels of nitrosylated PDI as compared with those of healthy controls (204). PDI nitrosylation prevents PDI-mediated ER stress reduction and allows protein misfolding (204).

Molecular chaperone Hsp90 is a target of *S*-nitrosylation, with critical cysteine residue susceptibility in the region of the C-terminal domain that interacts with endothelial nitric oxide synthase (eNOS). Hsp90 is another protein associated with AD that undergoes nitrosylation (107). Hsp90 ATPase activity has a general positive effect on eNOS activity. Both, however, are inhibited by *S*-nitrosylation. *S*-Nitrosylation may therefore functionally regulate the general activities of Hsp90 and provide a feedback mechanism for limiting eNOS activation. Consistent with this, postmortem analysis of brain samples from patients with AD revealed increased levels of Hsp90. *S*-Nitrosylation of Hsp90 abolishes ATPase activity that is necessary for its chaperone function; thus, inactivation of Hsp90 may allow accumulation of tau and amyloid- β aggregates in the AD brain (108).

It is generally recognized that mitochondria continuously undergo two opposing processes, fission and fusion. The disruption of this dynamic equilibrium may herald cell injury or death and may contribute to developmental and neurodegenerative disorders. Nitric oxide produced in response to β -amyloid protein has been shown to trigger mitochondrial fission, synaptic loss, and neuronal damage, in part through *S*-nitrosylation of dynamin-related protein 1 (forming SNO-Drp1) (42). Preventing nitrosylation of Drp1 by cysteine mutation abrogated these neurotoxic events. Accordingly, SNO-Drp1 is increased in brains of AD patients and may thus contribute to the pathogenesis of neurodegeneration (42).

Parkinson's and Huntington's diseases

Studies have provided a considerable link between NO and other prevalent neurodegenerative diseases, such as PD and HD. PD, whose cardinal features include tremor, slowness of movement, stiffness, and poor balance, is attributed to a profound deficit in dopamine that follows the loss of dopaminergic neurons in the *substantia nigra pars compacta* and dopaminergic nerve terminals in the *striatum* (206). Although the mechanisms leading to PD are still uncertain, a large amount of experimental evidence implicates oxidative and nitrosative stress as crucial factors in the pathogenesis of PD (31). Considerable insights into the pathogenesis of PD have been achieved by use of the neurotoxin 1-methyl-4-phenyl-1,2,3,6-tetrahydropyridine (MPTP), which is commonly used to induce an experimental model of PD (167). Excessive free radical formation or antioxidant deficiency and the resulting oxidative stress are all mechanisms involved in MPTP neurotoxicity (167). Animal models of MPTP-induced neurotoxicity have shown that inhibition of NOS slows the progression of disease pathology (213). MPTP is a neurotoxin that inhibits complex I of the mitochondrial respiratory chain and mimics the symptoms of PD by killing substantia nigra neurons (31). Of note, 7-nitroindazole (7-NI), a selective inhibitor of nNOS, blocks MPTP-mediated decrease in striatal dopamine levels as well as neuronal death in mice (210) and baboons (70). 7-NI also protects against motor deficits and cognitive decline in the MPTP baboon model (108). In addition to the role of nNOS in MPTP-induced neurotoxicity, iNOS appears to play a role. MPTP treatment of mice causes massive gliosis, a proliferation of glial cells, and upregulation of iNOS, and iNOS-deficient mice are more resistant to MPTP (108).

In HD, another age-related neurodegenerative disorder that often gives rise to dementia, evidence of oxidative and nitrosative damage exists in the basal ganglia, as was recently reviewed (17). It was shown that matrix metalloproteinase 9 (MMP9), which causes neuronal apoptosis, is S-nitrosylated by NO that is derived from the endogenous nitrosothiol S-nitrosocysteine (32). Matrix metalloproteinases are involved in the pathogenesis of acute and chronic neurodegenerative disorders such as stroke, AD, HIV-associated dementia, and multiple sclerosis (11). A similar mechanism, proposed to occur in PD (46, 212), independently demonstrated that S-nitrosocysteine-derived NO is able to nitrosylate parkin, an E3 ubiquitin ligase. Mutation in parkin is known to cause autosomal recessive juvenile parkinsonism. Nitrosylation of cysteine residues of parkin initially increases but later decreases the E3 ubiquitin ligase activity of this protein, and thereby reduces its protective function. NO has also been shown to S-nitrosylate GAPDH, thereby reducing its activity and enabling GAPDH to bind to another E3 ubiquitin ligase, Siah1. The GAPDH-Siah1 complex then translocates into the nucleus to induce apoptosis. As a direct consequence of this finding, the identification of a new mechanism of action for selegiline, a drug already used to treat patients with PD, has emerged. At nanomolar concentrations, selegiline prevents S-nitrosylation of GAPDH, thereby blocking its interaction with Siah1 and the further induction of apoptosis. The neuroprotective effect selegiline is similar to that of TCH346, a derivative with no monoamine oxidase type B inhibitory activity (85). In addition, peroxiredoxin 2 (PRX2), a member of a

family of abundant antioxidants most commonly found in neurons that reduce intracellular peroxides, becomes S-nitrosylated in a reaction with NO (108). S-Nitrosylation of PRX2 prevents its reaction with peroxides and inhibits its enzymatic activity and protective function against oxidative stress. Studies of human postmortem brains revealed an increase in S-nitrosylated PRX2 in PD patients (108).

Nitrated proteins identified in disease

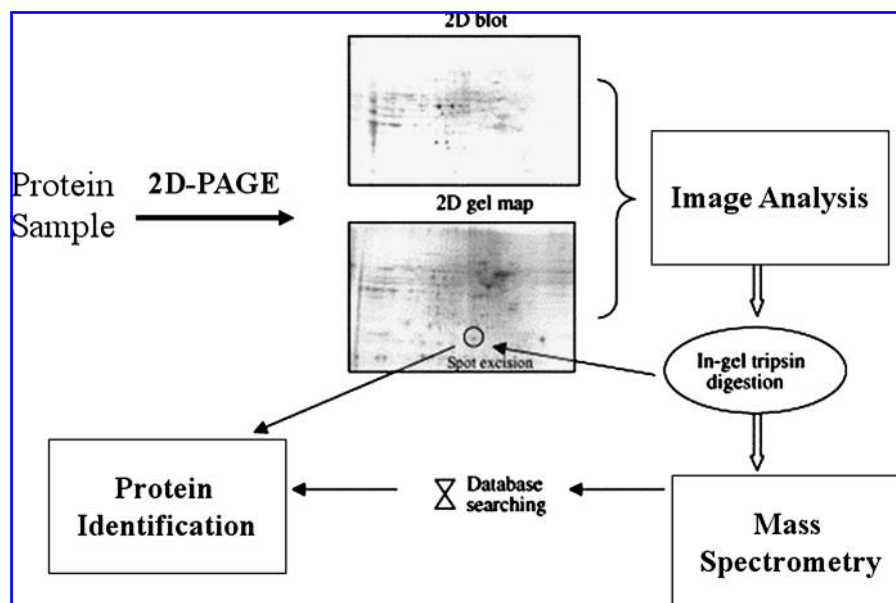
Proteins are particularly susceptible to nitration at tyrosine, cysteine, methionine, tryptophan, and histidine residues and at transition metal redox centers (158). Tyrosine is the most studied nitrated amino acid and has been identified in >50 human diseases and >80 animal models (158). The first protein identified to be nitrated *in vivo* was mitochondrial Mn-SOD, and nitration of a single tyrosine residue (Tyr-34) completely inactivates the enzyme (123). Nitrated Mn-SOD has been observed in multiple human disease conditions, including rejected kidney allografts, the cerebrospinal fluid of amyotrophic lateral sclerosis, AD and PD patients, diabetic heart tissue, and vascular aging (211). Protein tyrosine nitration in the circulatory and central nervous systems has been implicated in multiple disease conditions (195, 196). Creatine kinase and the sarcoplasmic reticulum Ca^{2+} ATPase (SERCA) are inactivated by tyrosine nitration (114). Creatine kinase is the energetic regulator of cardiomyocyte contraction, and the SERCA is a major regulator of Ca^{2+} concentration in muscle cells. Inactivation of these proteins has major implications for the development or cause (or both) of cardiovascular disease (157). Voltage-gated K^{+} channels are also nitrated in coronary endothelium and potentially lead to the impaired coronary flow observed in cardiac disease (115). In addition, α -actin, desmin, and myosin heavy chain, all proteins involved in contraction, are nitrated (14).

Tyrosine nitration has been studied extensively in neurodegenerative diseases. In PD, α -synuclein nitration leads to oligomerization and subsequent Lewy body formation, the major pathologic hallmark of PD (5). Tyrosine-hydroxylase, the enzyme that catalyzes the rate-limiting step in dopamine synthesis, also is nitrated in PD (161). In amyotrophic lateral sclerosis (ALS), neurofilament L is nitrated and may be involved in the aberrant motor neuron function characteristic of ALS (53). In AD, studies have shown that 3,3'-dinitrotyrosine and 3-nitrotyrosine levels in the hippocampus, neocortex, and ventricular cerebrospinal fluid are five- to eightfold higher compared with age-matched controls (113). Redox proteomics techniques (Fig. 8) have been used to identify 10 proteins that show increased specific nitrotyrosine immunoreactivity in the brains of patients with AD. Specific proteins found to be nitrated in AD include α -enolase, triose-phosphate isomerase, neuropolypeptide h3, β -actin, L-lactate dehydrogenase, carbonic anhydrase II, glyceraldehyde 3-phosphate dehydrogenase (GAPDH), ATP synthase α chain, voltage-dependent anion channel protein 1, and γ -enolase (195).

Mild cognitive impairment (MCI), arguably the earliest form of AD, has several proteins that are nitrated, including α -enolase, aldolase, GAPDH, malate dehydrogenase, glucose-regulated protein (GRP) precursor, heat-shock protein 70, glutathione S-transferase 3 (GST3), multidrug-resistance protein 3 (MRP3), peroxiredoxin, dihydropyrimidase like-2

FIG. 8. Redox proteomics and the identification of nitrated proteins.

Redox proteomics has the potential of detecting disease markers and identifying potential targets for drug therapy in neurodegenerative disorders. Redox proteomics involves the separation of brain proteins followed by detection, usually immunochemically, of oxidatively modified proteins, either from a two-dimensional Western blot or from column eluents. Subsequent mass spectrometric analysis of tryptic digests and database searching leads to protein identification. The identified nitrated proteins play roles in different cellular functions that are directly or indirectly linked to pathophysiologic conditions.



(DRP2), fascin 1, and 14-3-3 γ (196). α -Enolase was previously identified as specifically oxidized in AD brain (39). Taken together with the increased nitration of the metabolic enzymes triosephosphate isomerase, aldolase, and malate dehydrogenase, these results indicate a possible mechanism to explain the altered glucose tolerance and metabolism exhibited in AD (197). In addition to metabolism, GAPDH plays a role as an NO sensor (45). Neuropolypeptide h3, also known as phosphatidylethanolamine-binding protein (PEBP), hippocampal cholinergic neurostimulating peptide (HCNP), and raf-kinase inhibitor protein (RKIP), has a variety of functions in the brain, including upregulating the production of acetylcholine transferase in cholinergic neurons after NMDA-receptor activation (151). Acetylcholine transferase activity is decreased in AD, and cholinergic deficits are prominent in AD brain (188). Nitration of neuropolypeptide h3 may contribute to the decline in cognitive function due to lack of neurotrophic action on cholinergic neurons of the hippocampus and basal forebrain.

Carbonic anhydrase II is critical for maintenance of pH and control of carbon dioxide levels, and its activity is altered in AD (197). ATP synthase α chain is important in energy metabolism, and voltage-dependent anion channel protein 1 is involved in the mitochondrial permeability transition pore, with consequent apoptotic considerations, as well as in mitochondrial calcium ion homeostasis.

GRP precursor and Hsp70 are both chaperone proteins that aid in the proper folding of molecular targets and escort targets to the proteasome for degradation if proper folding is not achieved. MRP3 and GST3 are major detoxification enzymes that pump xenobiotics and glutathione-conjugated proteins out of the cell. The nitration of GRP precursor, Hsp70, MRP3, and GST3 proteins conceivably inhibits function and may contribute to the formation of the senile plaques observed in AD. DRP-2 is involved in axonal outgrowth and pathfinding to neighboring neurons. Fascin-1 is a marker of dendritic functionality, and the addition of fascin-1 has shown to be protective against oxidative stress (153). The nitration of

DRP-2 and fascin-1 may contribute to the shortened dendritic length observed in AD (155).

In rodent models of traumatic brain injury (TBI), a condition in which sudden trauma and secondary injury cause brain damage, nitrated tyrosine residues were found on several proteins (169). These proteins include synapsin 1, γ -enolase, guanosine diphosphate dissociation inhibitor 1 (GDP1), phosphoglycerate mutase (PGM), Hsp70, and ATP synthase (169). Synapsin 1 aids in the maintenance of synaptic vesicle reserves for the release of neurotransmitters (152). Nitrated synapsin 1 correlates with the reduced neurotransmission and synaptic plasticity that patients with TBI exhibit (184). GDP1 inhibits the exchange of GDP for GTP, a process necessary for glycogen breakdown to form glucose. Glycogen levels are increased in TBI brain compared with controls, a downstream effect impaired by GDP1 activity (154). Similar to cysteine nitration by peroxynitrite, which inactivates numerous proteins involved in energy production, including GAPDH, creatine kinase, and complexes I, II, III, and V of the electron-transport chain, several of these proteins are susceptible to tyrosine nitration, as described earlier, making metabolic enzymes particularly vulnerable to oxidative modification by peroxynitrite (108).

Methionine is directly oxidized by peroxynitrite predominantly to form methionine sulfoxide, but ethylene and dimethylsulfide are formed in some instances (4). Methionine oxidation inactivates glutamine synthetase, an enzyme responsible for glutamine synthesis. Glutamine is a critical amino acid for immune system activity and is a factor necessary for macrophage-mediated phagocytosis (179). Methionine oxidation is reversible and catalyzed by methionine sulfoxide reductase (MsrA). MsrA reduces methionine sulfoxide back to methionine at the expense of NAD(P)H. MsrA uses the thioredoxin/thioredoxin reductase endogenous antioxidant system to regenerate the catalytic active site (104). Decreased methionine reductase levels have been associated with the onset of AD (192, 193). Peroxynitrite also modifies tryptophan and histidine, but the implications of the nitration

of these amino acids are less clear and are the subject of current investigation (4).

Conclusions

As discussed in this review, traditional views of nitric oxide (NO) biology have been based around four key principles: (a) NO synthases [endothelial NO synthase (eNOS), neuronal NOS(nNOS), and inducible NOS (iNOS)] have unique and independent physiologic functions; (b) NO diffuses freely in biologic systems to carry out the tasks of each isoform; (c) the physiologic actions of NO are mediated primarily by cGMP, through the binding of NO to the heme group in soluble guanylate cyclase (sGC); and (d) the cytotoxic *versus* cytoprotective properties of NO are the consequence of the specific and respectively nonspecific redox chemistry of NO nitrosylation. Here we highlighted the Janus nature of NO. Depending on a number of factors, NO can be neuroprotective or neurotoxic. Clearly, additional molecular understanding of the roles of NO and their sequelae in cellular physiology, CNS injury, and CNS neuroprotection is needed. Newer, more powerful proteomics methods could be used to identify nitrated proteins that are not easily separated by using 2D gel electrophoretic methods. Methods of identifying low-abundance signaling proteins that are regulated or damaged by nitration will provide new insights into these Janus roles of NO. In addition, the possibility of counteracting NO/RNS neurotoxic effects by modulating the gene expression of endogenous enzymes and chaperones, which play key roles in defense against cellular injuries, by nutritional products or pharmacologic compounds, represents an innovative approach to therapeutic intervention in neurodegenerative disorders. The future of NO research in the CNS is bright, and much remains to be learned. Given the significant increase in the average age of the population in most Western and developing countries as health care improves, and the fact that aging is the single most important risk factor for age-related neurodegenerative disorders like AD, this is an exciting time to be involved in research aimed at deciphering the Janus roles of NO in the CNS.

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References

1. Abdul HM, Calabrese V, Calvani M, and Butterfield AD. Acetyl-L-carnitine-induced up-regulation of heat shock proteins protects cortical neurons against amyloid-beta peptide 1-42-mediated oxidative stress and neurotoxicity: implications for Alzheimer's disease. *J Neurosci Res* 84: 398–408, 2006.
2. Abraham M, Gola J, Cometto-Muniz E, and Cain W. The solvation properties of nitric oxide. *J Chem Soc Perkin Trans 2*: 2067–2070, 2000.
3. Aluise CD, St Clair D, Vore M, and Butterfield DA. In vivo amelioration of Adriamycin induced oxidative stress in plasma by gamma-glutamylcysteine ethyl ester (GCEE). *Cancer Lett* 282: 25–29, 2009.
4. Alvarez B and Radi R. Peroxynitrite reactivity with amino acids and proteins. *Amino Acids* 25: 295–311, 2003.
5. Andrekopoulou C, Zhang H, Joseph J, Kalivendi S, and Kalyanaraman B. Bicarbonate enhances alpha-synuclein oligomerization and nitration: intermediacy of carbonate radical anion and nitrogen dioxide radical. *Biochem J* 378: 435–447, 2004.
6. Baker PR, Schopfer FJ, Sweeney S, and Freeman BA. Red cell membrane and plasma linoleic acid nitration products: synthesis, clinical identification, and quantitation. *Proc Natl Acad Sci U S A* 101: 11577–11582, 2004.
7. Baker PRS, Schopfer FJ, O'Donnell VB, and Freeman BA. Convergence of nitric oxide and lipid signaling: anti-inflammatory nitro-fatty acids. *Free Radic Biol Med* (epub 2009).
8. Balogun E, Hoque M, Gong P, Killeen E, Green CJ, Foresti R, Alam J, and Motterlini R. Curcumin activates the haem oxygenase-1 gene via regulation of Nrf2 and the antioxidant-responsive element. *Biochem J* 371: 887–895, 2003.
9. Barone E, Trombino S, Cassano R, Sgambato A, De Paola B, Di Stasio E, Picci N, Preziosi P, and Mancuso C. Characterization of the S-denitrosylating activity of bilirubin. *J Cell Mol Med* (Epub 2009).
10. Beckman JS, Beckman TW, Chem J, Marshall PA, and Freeman BA. Apparent hydroxyl radical production by peroxynitrite: implications for endothelial injury from nitric oxide and superoxide. *Proc Natl Acad Sci U S A* 87: 1620–1624, 1990.
11. Benesová Y, Vasku A, Novotná H, Litzman J, Stourac P, Beránek M, Kadanka Z, and Bednarík J. Matrix metalloproteinase-9 and matrix metalloproteinase-2 as biomarkers of various courses in multiple sclerosis. *Mult Scler* 15: 316–322, 2009.
12. Benhar M, Forrester MT, Hess DT, and Stamler JS. Regulated protein denitrosylation by cytosolic and mitochondrial thioredoxins. *Science* 320, 1050–1054, 2008.
13. Bon CL and Garthwaite J. On the role of nitric oxide in hippocampal long-term potentiation. *J Neurosci* 23: 1941–1948, 2003.
14. Borbely A, Toth A, Edes I, Virag L, Papp JG, Varro A, Paulus WJ, van der Velden J, Stienen GJ, and Papp Z. Peroxynitrite-induced alpha-actinin nitration and contractile alterations in isolated human myocardial cells. *Cardiovasc Res* 67: 225–233, 2005.
15. Boulton CL, Southam E, and Garthwaite J. Nitric oxide-dependent long-term potentiation is blocked by a specific inhibitor of soluble guanylyl cyclase. *Neuroscience* 69: 699–703, 1995.
16. Brown GC and Bal-Price A. Inflammatory neurodegeneration mediated by nitric oxide, glutamate, and mitochondria. *Mol Neurobiol* 27: 325–355, 2003.
17. Browne SE and Beal MF. Oxidative damage in Huntington's disease pathogenesis. *Antioxid Redox Signal* 8: 2061–2073, 2006.
18. Buckley BJ, Marshall ZM, and Whorton AR. Nitric oxide stimulates Nrf2 nuclear translocation in vascular endothelium. *Biochem Biophys Res Commun* 30: 973–979, 2003.
19. Buckley BJ, Li S, and Whorton AR. Keap1 modification and nuclear accumulation in response to S-nitrosocysteine. *Free Radic Biol Med* 44: 692–698, 2008.
20. Butterfield DA and Sultana R. Identification of 3-nitrotyrosine-modified brain proteins by redox proteomics. *Methods Enzymol* 440: 295–308, 2008.

21. Calabrese V, Copani A, Testa D, Ravagna A, Spadaro F, Tendi E, Nicoletti V, and Giuffrida Stella AM. Nitric oxide synthase induction in astroglial cell cultures: effect on heat shock protein 70 synthesis and oxidant/antioxidant balance. *J Neurosci Res* 60: 613–622, 2000.
22. Calabrese V, Scapagnini G, Ravagna A, Bella R, Foresti R, Bates TE, Giuffrida Stella AM, and Pennisi G. Nitric oxide synthase is present in the cerebrospinal fluid of patients with active multiple sclerosis and is associated with increases in cerebrospinal fluid protein nitrotyrosine and S-nitrosothiols and with changes in glutathione levels. *J Neurosci Res* 70: 580–587, 2002.
23. Calabrese V, Boyd-Kimball D, Scapagnini G, and Butterfield DA. Nitric oxide and cellular stress response in brain aging and neurodegenerative disorders: the role of vitagenes. *In Vivo* 18: 245–267, 2004.
24. Calabrese V, Giuffrida Stella AM, Butterfield DA, and Scapagnini G. Redox regulation in neurodegeneration and longevity: role of the heme oxygenase and HSP70 systems in brain stress tolerance. *Antioxid Redox Signal* 6: 895–913, 2004.
25. Calabrese V, Colombrita C, Guagliano E, Sapienza M, Ravagna A, Tomaselli G, Cardile V, Scapagnini G, Butterfield DA, Giuffrida Stella AM, and Rizzarelli E. Protective effect of carnosine during nitrosative stress in astroglial cell cultures. *Neurochem Res* 30: 797–807, 2005.
26. Calabrese V, Ravagna A, Colombrita C, Guagliano E, Scapagnini G, Calvani M, Butterfield DA, and Giuffrida Stella AM. Acetylcarnitine induces heme oxygenase in rat astrocytes and protects against oxidative stress: involvement of the transcription factor Nrf2. *J Neurosci Res* 79: 509–521, 2005.
27. Calabrese V, Butterfield DA, Scapagnini G, Stella AM, and Maines MD. Redox regulation of heat shock protein expression by signaling involving nitric oxide and carbon monoxide: relevance to brain aging, neurodegenerative disorders, and longevity. *Antioxid Redox Signal* 8: 444–477, 2006.
28. Calabrese V, Sultana R, Scapagnini G, Guagliano E, Sapienza M, Bella R, Kanski J, Pennisi G, Mancuso C, Stella AM, and Butterfield DA. Nitrosative stress, cellular stress response, and thiol homeostasis in patients with Alzheimer's disease. *Antioxid Redox Signal* 8: 1975–1986, 2006.
29. Calabrese V, Colombrita C, Scapagnini G, Calvani M, Giuffrida Stella AM, and Butterfield DA. Acetylcarnitine and cellular stress response: role in nutritional redox homeostasis and regulation of longevity genes. *J Nutr Biochem* 17: 73–88, 2006.
30. Calabrese V, Calvani M, and Butterfield DA. Acetyl-L-carnitine-induced up-regulation of heat shock proteins protects cortical neurons against amyloid-beta peptide 1-42-mediated oxidative stress and neurotoxicity: implications for Alzheimer's disease. *J Neurosci Res* 84: 398–408, 2006.
31. Calabrese V, Guagliano E, Sapienza M, Panebianco M, Calafato S, Puleo E, Pennisi G, Mancuso C, Butterfield DA, and Stella AG. Redox regulation of cellular stress response in aging and neurodegenerative disorders: role of vitagenes. *Neurochem Res* 32: 757–773, 2007.
32. Calabrese V, Mancuso C, Calvani M, Rizzarelli E, Butterfield DA, and Giuffrida Stella AM. Nitric oxide in the central nervous system: neuroprotection versus neurotoxicity. *Nat Rev Neurosci* 8: 766–775, 2007.
33. Calabrese V, Butterfield DA, and Stella AM. Aging and oxidative stress response in the CNS. In: *Development and aging changes in the nervous system: handbook of neurochemistry and molecular neurobiology*, Lajtha A, Perez-Polo JR, and Rossner S, eds. 3rd ed, pp 128–234, 2008, NY: Springer.
34. Calabrese V, Mancuso C, Cornelius C, Calafato M, Ventimiglia B, Butterfield DA, Dinkova-Kostova AT, and Rizzarelli E. Reactive nitrogen species and cellular stress tolerance in aging and neurodegeneration: role of vitagenes. In: *Free radical pathophysiology*, Alvarez S, and Evelson P, eds. pp. 345–367, Kerala, India: Transworld Research Network, 2008.
35. Calabrese V, Cornelius C, Mancuso C, Barone E, Calafato S, Bates T, Rizzarelli E, and Dinkova-Kostova AT. Vitagenes, dietary antioxidants and neuroprotection in neurodegenerative diseases. *Front Biosci* 14: 376–397, 2009.
36. Cardone MH, Roy N, Stennicke HR, Salvesen GS, Franke TF, Stanbridge E, Frisch S, and Reed JC. Regulation of cell death protease caspase-9 by phosphorylation. *Science* 282: 1318–1321, 1998.
37. Carreras MC and Poderoso JJ. Mitochondrial nitric oxide in the signaling of cell integrated responses. *Am J Physiol Cell Physiol* 292: 1569–1580, 2007.
38. Carreras MC, Franco MC, Finocchietto PV, Converso DP, Antico Arciuch VG, Holod S, Peralta JG, and Poderoso JJ. The biological significance of mtNOS modulation. *Front Biosci* 12: 1041–1048, 2007.
39. Castegna A, Aksenov M, Thongboonkerd V, Klein JB, Pierce WM, Booze R, Markesbery WR, and Butterfield DA. Proteomic identification of oxidatively modified proteins in Alzheimer's disease brain, part II: dihydropyrimidinase-related protein 2, alpha-enolase and heat shock cognate 71. *J Neurochem* 82: 1524–1532, 2002.
40. Chang AY, Chan JY, Cheng HL, Tsai CY, and Chan SH. Hypoxia-inducible factor-1/heme oxygenase-1 cascade as upstream signals in the pro-life role of heat shock protein 70 at rostral ventrolateral medulla during experimental brain stem death. *Shock* (Epub 2009).
41. Chien WL, Liang KC, Teng CM, Kuo SC, Lee FY, and Fu WM. Enhancement of long-term potentiation by a potent nitric oxide-guanylyl cyclase activator, 3-(5-hydroxymethyl-2-furyl)-1-benzyl-indazole. *Mol Pharmacol* 63: 1322–1328, 2003.
42. Cho DH, Nakamura T, Fang J, Cieplak P, Godzik A, Gu Z, and Lipton SA. S-Nitrosylation of Drp1 mediates beta-amyloid-related mitochondrial fission and neuronal injury. *Science* 324: 102–105, 2009.
43. Choi SH, Aid S, and Bosetti F. The distinct roles of cyclooxygenase-1 and -2 in neuroinflammation: implications for translational research. *Trends Pharmacol Sci* 30: 174–181, 2009.
44. Choi YB and Lipton SA. Redox modulation of the NMDA receptor. *Cell Mol Life Sci* 57: 1535–1541, 2000.
45. Chuang DM, Hough C, and Senatorov VV. Glyceraldehyde-3-phosphate dehydrogenase, apoptosis, and neurodegenerative diseases. *Annu Rev Pharmacol Toxicol* 45: 269–290, 2005.
46. Chung KK, Thomas B, Li X, Pletnikova O, Troncoso JC, Marsh L, Dawson VL, and Dawson TM. S-Nitrosylation of parkin regulates ubiquitination and compromises parkin's protective function. *Science* 304: 1328–1331, 2004.
47. Ciani E, Guidi S, Bartesaghi R, and Contestabile A. Nitric oxide regulates cGMP-dependent cAMP-responsive element binding protein phosphorylation and Bcl-2 expression in cerebellar neurons: implication for a survival role of nitric oxide. *J Neurochem* 82: 1282–1289, 2002.
48. Ciani E, Guidi S, Della Valle G, Perini G, Bartesaghi R, and Contestabile A. Nitric oxide protects neuroblastoma cells

- from apoptosis induced by serum deprivation through cAMP-response element-binding protein (CREB) activation. *J Biol Chem* 277: 49896–49902, 2002.
49. Ciani E, Virgili M, and Contestabile A. Akt pathway mediates a cGMP-dependent survival role of nitric oxide in cerebellar granule neurones. *J Neurochem* 81: 218–228, 2002.
 50. Coffey MJ, Phare SM, and Peters-Golden M. Interaction between nitric oxide, reactive oxygen intermediates, and peroxynitrite in the regulation of 5-lipoxygenase metabolism. *Biochim Biophys Acta* 1584: 81–90, 2002.
 51. Contestabile A and Ciani E. Role of nitric oxide in the regulation of neuronal proliferation, survival and differentiation. *Neurochem Int* 45: 903–914, 2004.
 52. Contestabile A. Regulation of transcription factors by nitric oxide in neurons and in neural-derived tumor cells. *Prog Neurobiol* 84: 317–328, 2008.
 53. Crow JP, Ye YZ, Strong M, Kirk M, Barnes S, and Beckman JS. Superoxide dismutase catalyzes nitration of tyrosines by peroxynitrite in the rod and head domains of neurofilament-L. *J Neurochem* 69: 1945–1953, 1997.
 54. Cullinan SB, Gordan JD, Jin J, Harper JW, and Diehl JA. The Keap1-BTB protein is an adaptor that bridges Nrf2 to a Cul3-based E3 ligase: oxidative stress sensing by a Cul3-Keap1 ligase. *Mol Cell Biol* 24: 8477–8486, 2004.
 55. Cumming RC and Schubert D. Amyloid-beta induces disulfide bonding and aggregation of GAPDH in Alzheimer's disease. *FASEB J* 19: 2060–2062, 2005.
 56. Datta SR, Dudek H, Tao X, Masters S, Fu H, Gotoh Y, and Greenberg ME. Akt phosphorylation of BAD couples survival signals to the cell-intrinsic death machinery. *Cell* 91: 231–241, 1997.
 57. Dawson VL and Dawson TM. Nitric oxide in neurodegeneration. *Prog Brain Res* 118: 215–229, 1998.
 58. Dawson TM and Snyder SH. Gases as biological messengers: nitric oxide and carbon monoxide in the brain. *J Neurosci* 14: 5147–5159, 1994.
 59. De Palma C, Falcone S, Panzeri C, Radice S, Bassi MT, and Clementi E. Endothelial nitric oxide synthase overexpression by neuronal cells in neurodegeneration: a link between inflammation and neuroprotection. *J Neurochem* 106: 193–204, 2008.
 60. Dedon PC and Tannenbaum SR. Reactive nitrogen species in the chemical biology of inflammation. *Arch Biochem Biophys* 423: 12–22, 2004.
 61. Delgado-Esteban M, Martin-Zanca D, Andres-Martin L, Almeida A, and Bolaños JP. Inhibition of PTEN by peroxynitrite activates the phosphoinositide-3-kinase/Akt neuroprotective signaling pathway. *J Neurochem* 102: 194–205, 2007.
 62. Denicola A, Freeman BA, Trujillo M, and Radi R. Peroxynitrite reaction with carbon dioxide/bicarbonate: kinetics and influence on peroxynitrite mediated oxidations. *Arch Biochem Biophys* 333: 49–58, 1996.
 63. Dhakshinamoorthy S and Porter AG. Nitric oxide-induced transcriptional up-regulation of protective genes by Nrf2 via the antioxidant response element counteracts apoptosis of neuroblastoma cells. *J Biol Chem* 279: 20096–20107, 2004.
 64. Ding H and Demple B. Direct nitric oxide signal transduction via nitrosylation of iron-sulfur centers in the SoxR transcription activator. *Proc Natl Acad Sci U S A* 97: 5146–5150, 2000.
 65. Dinkova-Kostova AT, Holtzclaw WD, Cole RN, Itoh K, Wakabayashi N, Katoh Y, Yamamoto M, and Talalay P. Direct evidence that sulfhydryl groups of Keap1 are the sensors regulating induction of phase 2 enzymes that protect against carcinogens and oxidants. *Proc Natl Acad Sci U S A* 99: 11908–11913, 2002.
 66. Dinkova-Kostova AT, Holtzclaw WD, and Kensler TW. The role of Keap1 in cellular protective responses. *Chem Res Toxicol* 18: 1779–1791, 2005.
 67. Dinkova-Kostova AT and Talalay P. Direct and indirect antioxidant properties of inducers of cytoprotective proteins. *Mol Nutr Food Res* 52: 128–138, 2008.
 68. Duman RS, Terwilliger RZ, and Nestler EJ. Alterations in nitric oxide-stimulated endogenous ADP-ribosylation associated with long-term potentiation in rat hippocampus. *J Neurochem* 61: 1542–1545, 1993.
 69. Elfering SL, Sarkela TM, and Giulivi C. Biochemistry of mitochondrial nitric-oxide synthase. *J Biol Chem* 277: 38079–38086, 2002.
 70. Ferrante RJ, Hantraye P, Brouillet E, and Beal MF. Increased nitrotyrosine immunoreactivity in substantia nigra neurons in MPTP treated baboons is blocked by inhibition of neuronal nitric oxide synthase. *Brain Res* 823: 177–182, 1999.
 71. Forman HJ, Fukuto JM, and Torres M. Redox signaling: thiol chemistry defines which reactive oxygen and nitrogen species can act as second messengers. *Am J Physiol Cell Physiol* 287: 246–256, 2004.
 72. Foster MW, McMahon TJ, and Stamler JS. S-nitrosylation in health and disease. *Trends Mol Med* 9: 160–168, 2003.
 73. Franco MC, Antico Arciuch VG, Peralta JG, Galli S, Levisman D, Lopez LM, Romorini L, Poderoso JJ, and Carreras MC. Hypothyroid phenotype is contributed by mitochondrial complex I inactivation due to translocated neuronal nitric oxide synthase. *J Biol Chem* 281: 4779–4786, 2006.
 74. Friling RS, Bensimon A, Tichauer Y, and Daniel V. Xenobiotic-inducible expression of murine glutathione S-transferase Ya subunit gene is controlled by an electrophile-responsive element. *Proc Natl Acad Sci U S A* 87: 6258–6262, 1990.
 75. Furchgott RF and Zawadzki JV. 1980. The obligatory role of endothelial cells in the relaxation of arterial smooth muscle by acetylcholine. *Nature* 288: 373–376, 1980.
 76. Furukawa M and Xiong Y. BTB protein Keap1 targets antioxidant transcription factor Nrf2 for ubiquitination by the Cullin 3-Roc1 ligase. *Mol Cell Biol* 25: 162–171, 2005.
 77. Gao J, Kashfi K, Liu X, Rigas B. NO-donating aspirin induces phase II enzymes in vitro and in vivo. *Carcinogenesis* 27: 803–810, 2006.
 78. Garthwaite J, Charles SL, and Chess-Williams R. Endothelium-derived relaxing factor release on activation of NMDA receptors suggests role as intercellular messenger in the brain. *Nature* 336: 385–388, 1988.
 79. Gaston BM, Carver J, Doctor A, and Palmer LA. S-Nitrosylation signaling in cell biology. *Mol Interv* 3: 253–263, 2003.
 80. Gonzalez-Cadavid NF and Rajfer J. The pleiotropic effects of inducible nitric oxide synthase (iNOS) on the physiology and pathology of penile erection. *Curr Pharm Des* 11: 4041–4046, 2005.
 81. Gow AJ, Buerk DG, and Ischiropoulos H. A novel reaction mechanism for the formation of S-nitrosothiol in vivo. *J Biol Chem* 272: 2841–2845, 1997.
 82. Gow AJ and Stamler JS. Reactions between nitric oxide and haemoglobin under physiological conditions. *Nature* 391: 169–173, 1998.
 83. Gu Z, Kaul M, Yan B, Kridel SJ, Cui J, Strongin A, Smith JW, Liddington RC, and Lipton SA. S-Nitrosylation of matrix

- metalloproteinases: signaling pathway to neuronal cell death. *Science* 297: 1186–1190, 2002.
84. Guix FX, Uribealago I, Coma M, and Munoz FJ The physiology and pathophysiology of nitric oxide in the brain. *Prog Neurobiol* 76: 126–152, 2005.
 85. Hara MR, Thomas B, Cascio MB, Bae BI, Hester LD, Dawson VL, Dawson TM, Sawa A, and Snyder SH. Neuroprotection by pharmacologic blockade of the GAPDH death cascade. *Proc Natl Acad Sci U S A* 103: 3887–3889, 2006.
 86. Haridas V, Kim SO, Nishimura G, Hausladen A, Stamler JS, and Gutterman JU. Avicinylation (thioesterification): a protein modification that can regulate the response to oxidative and nitrosative stress. *Proc Natl Acad Sci U S A* 102: 10088–10093, 2005.
 87. Hausladen A, Privalle CT, Keng T, DeAngelo J, and Stamler JS. Nitrosative stress: activation of the transcription factor OxyR. *Cell* 86: 719–729, 1996.
 88. Hausladen A, Gow AJ, and Stamler JS. Nitrosative stress: metabolic pathway involving the flavohemoglobin. *Proc Natl Acad Sci U S A* 95: 14100–14105, 1998.
 89. Hayes JD and McMahon M. NRF2 and KEAP1 mutations: permanent activation of an adaptive response in cancer. *Trends Biochem Sci* 34: 176–188, 2009.
 90. Heales SJ, Bolaños JP, Stewart VC, Brookes PS, Land JM, and Clark JB. Nitric oxide, mitochondria and neurological disease. *Biochim Biophys Acta* 1410: 215–228, 1999.
 91. Herold S, Exner M, and Boccini F. The mechanism of the peroxynitrite-mediated oxidation of myoglobin in the absence and presence of carbon dioxide. *Chem Res Toxicol* 16: 390–402, 2003.
 92. Hess DT, Matsumoto A, Kim SO, Marshall HE, and Stamler JS. Protein S-nitrosylation: purview and parameters. *Nat Rev Mol Cell Biol* 6: 150–166, 2005.
 93. Ignarro LJ. Signal transduction mechanisms involving nitric oxide. *Biochem Pharmacol* 41: 485–490, 1991.
 94. Ischiropoulos H. Biological selectivity and functional aspects of protein tyrosine nitration. *Biochem Biophys Res Commun* 305: 776–783, 2003.
 95. Itoh K, Igarashi K, Hayashi N, Nishizawa M, and Yamamoto M. Cloning and characterization of a novel erythroid cell-derived CNC family transcription factor heterodimerizing with the small Maf family proteins. *Mol Cell Biol* 15: 4184–4193, 1995.
 96. Itoh K, Wakabayashi N, Katoh Y, Ishii T, Igarashi K, Engel JD, and Yamamoto M. Keap1 represses nuclear activation of antioxidant responsive elements by Nrf2 through binding to the amino-terminal Neh2 domain. *Genes Dev* 13: 76–86, 1999.
 97. Iwakiri Y, Satoh A, Chatterjee S, Toomre DK, Chalouni CM, Fulton D, Groszmann RJ, Shah VH, and Sessa WC. Nitric oxide synthase generates nitric oxide locally to regulate compartmentalized protein S-nitrosylation and protein trafficking. *Proc Natl Acad Sci U S A* 103: 19777–19782, 2006.
 98. Jaffrey SR, Erdjument-Bromage H, Ferris CD, Tempst P, and Snyder SH. Protein S-nitrosylation: a physiological signal for neuronal nitric oxide. *Nat Cell Biol* 3: 193–197, 2001.
 99. Janssen-Heininger YM, Mossman BT, Heintz NH, Forman HJ, Kalyanaraman B, Finkel T, Stamler JS, Rhee SG, and van der Vliet A. Redox-based regulation of signal transduction: principles, pitfalls, and promises. *Free Radic Biol Med* 45: 1–17, 2008.
 100. Kaehler ST, Singewald N, Sinner C, and Philippu A. Nitric oxide modulates the release of serotonin in the rat hypothalamus. *Brain Res* 835: 346–349, 1999.
 101. Kan WH, Hsu JT, Schwacha MG, Choudhry MA, Raju R, Bland KI, and Chaudry IH. Selective inhibition of iNOS attenuates trauma-hemorrhage/resuscitation-induced hepatic injury. *J Appl Physiol* 105: 1076–1082, 2008.
 102. Kedia GT, Uckert S, Jonas U, Kuczyk MA, and Burchardt M. The nitric oxide pathway in the human prostate: clinical implications in men with lower urinary tract symptoms. *World J Urol* 26: 603–639, 2008.
 103. Kensler TW, Wakabayashi N, and Biswal S. Cell survival responses to environmental stresses via the Keap1-Nrf2-ARE pathway. *Annu Rev Pharmacol Toxicol* 47: 89–116, 2007.
 104. Kim HY and Kim JR. Thioredoxin as a reducing agent for mammalian methionine sulfoxide reductases B lacking resolving cysteine. *Biochem Biophys Res Commun* 371: 490–494, 2008.
 105. Kim SO, Merchant K, Nudelman R, Beyer WF, Keng T, DeAngelo J, Hausladen A, and Stamler JS. OxyR: a molecular code for redox-related signaling. *Cell* 109: 383–396, 2002.
 106. Kitamura Y, Furukawa M, Matsuoka Y, Tooyama I, Kimura H, Nomura Y, and Taniguchi T. In vitro and in vivo induction of heme oxygenase-1 in rat glial cells: possible involvement of nitric oxide production from inducible nitric oxide synthase. *Glia* 22: 138–148, 1998.
 107. Knott AB and Bossy-Wetzel E. Impairing the mitochondrial fission and fusion balance: a new mechanism of neurodegeneration. *Ann N Y Acad Sci* 1147: 283–292, 2008.
 108. Knott AB and Bossy-Wetzel E. Nitric oxide in health and disease of the nervous system. *Antioxid Redox Signal* 11: 541–553, 2009.
 109. Kobayashi A, Kang MI, Okawa H, Ohtsuiji M, Zenke Y, Chiba T, Igarashi K, and Yamamoto M. Oxidative stress sensor Keap1 functions as an adaptor for Cul3-based E3 ligase to regulate proteasomal degradation of Nrf2. *Mol Cell Biol* 24: 7130–7139, 2004.
 110. Kobayashi M and Yamamoto M. Nrf2-Keap1 regulation of cellular defense mechanisms against electrophiles and reactive oxygen species. *Adv Enzyme Regul* 46: 113–140, 2006.
 111. Kone BC, Kuncewicz T, Zhang W, and Yu ZY. Protein interactions with nitric oxide synthases: controlling the right time, the right place, and the right amount of nitric oxide. *Am J Physiol Renal Physiol* 285: F178–F190, 2003.
 112. Koppenol WH, Moreno JJ, Pryor WA, Ischiropoulos H, and Beckman JS. Peroxynitrite, a cloaked oxidant formed by nitric oxide and superoxide. *Chem Res Toxicol* 5: 834–842, 1992.
 113. Korolainen MA and Pirttilä T. Cerebrospinal fluid, serum and plasma protein oxidation in Alzheimer's disease. *Acta Neurol Scand* 119: 32–38, 2009.
 114. Lee WH, Gounarides JS, Roos ES, and Wolin MS. Influence of peroxynitrite on energy metabolism and cardiac function in a rat ischemia-reperfusion model. *Am J Physiol Heart Circ Physiol* 285: H1385–H1395, 2003.
 115. Li H, Gutterman DD, Rusch NJ, Bubolz A, and Liu Y. Nitration and functional loss of voltage-gated K⁺ channels in rat coronary microvessels exposed to high glucose. *Diabetes* 53: 2436–2442, 2004.
 116. Lipton SA, Singel DJ, and Stamler JS. Nitric oxide in the central nervous system. *Prog Brain Res* 103: 359–364, 1994.
 117. Lipton SA, Choi YB, Takahashi H, Zhang D, Li W, Godzik A, and Bankston LA. Cysteine regulation of protein function: as exemplified by NMDA-receptor modulation. *Trends Neurosci* 25: 474–480, 2002.

118. Liu L and Stamler JS. NO: an inhibitor of cell death. *Cell Death Differ* 6: 937–942, 1999.
119. Liu XM, Peyton KJ, Ensenat D, Wang H, Hannink M, Alam J, and Durante W. Nitric oxide stimulates heme oxygenase-1 gene transcription via the Nrf2/ARE complex to promote vascular smooth muscle cell survival. *Cardiovasc Res* 75: 381–389, 2007.
120. Lonart G, Wang J, and Johnson KM. Nitric oxide induces neurotransmitter release from hippocampal slices. *Eur J Pharmacol* 220: 271–272, 1992.
121. Lorrain DS and Hull EM. Nitric oxide increases dopamine and serotonin release in the medial preoptic area. *Neuroreport* 5: 87–89, 1993.
122. Lymar SV and Hurst JK. Rapid reaction between peroxynitrite and carbon dioxide: implications for biological activity. *J Am Chem Soc* 117: 8867–8868, 1995.
123. MacMillan-Crow LA and Thompson JA. Tyrosine modifications and inactivation of active site manganese superoxide dismutase mutant (Y34F) by peroxynitrite. *Arch Biochem Biophys* 366: 82–88, 1999.
124. Mancuso C, Bates TE, Butterfield DA, Calafato S, Cornelius C, De Lorenzo A, Dinkova Kostova AT, and Calabrese V. Natural antioxidants in Alzheimer's disease. *Expert Opin Invest Drugs* 16: 1921–1931, 2007.
125. Mancuso C, Scapagini G, Currò D, Giuffrida Stella AM, De Marco C, Butterfield DA, and Calabrese V. Mitochondrial dysfunction, free radical generation and cellular stress response in neurodegenerative disorders. *Front Biosci* 12: 1107–1123, 2007.
126. Mannick JB, Hausladen A, Liu L, Hess DT, Zeng M, Miao QX, Kane LS, Gow AJ, and Stamler JS. Fas-induced caspase denitrosylation. *Science* 284: 651–654, 1999.
127. Mannick JB. Regulation of apoptosis by protein S-nitrosylation. *Amino Acids* 32: 523–526, 2007.
128. Mannick JB and Schonhoff CM. Measurement of protein S-nitrosylation during cell signalling. *Methods Enzymol* 440: 231–242, 2008.
129. Marshall HE and Stamler JS. Nitrosative stress-induced apoptosis through inhibition of NF-kappa B. *J Biol Chem* 277: 34223–34228, 2002.
130. Martínez MC and Andriantsitohaina R. Reactive nitrogen species: molecular mechanisms and potential significance in health and disease. *Antioxid Redox Signal* 11: 669–702, 2009.
131. McCann SM. The nitric oxide hypothesis of brain aging. *Exp Gerontol* 32: 431–440, 1997.
132. McMahon M, Itoh K, Yamamoto M, and Hayes JD. Keap1-dependent proteasomal degradation of transcription factor Nrf2 contributes to the negative regulation of antioxidant response element-driven gene expression. *J Biol Chem* 278: 21592–21600, 2003.
133. Melino G, Bernassola F, Knight RA, Corasaniti MT, Nistico G, and Finazzi-Agro AS. S-Nitrosylation regulates apoptosis. *Nature* 388: 432–433, 1997.
134. Mieyal JJ, Gallogly MM, Qanungo S, Sabens EA, and Shelton MD. Molecular mechanisms and clinical implications of reversible protein S-glutathionylation. *Antioxid Redox Signal* 10: 1941–1988, 2008.
135. Moi P, Chan K, Asunis I, Cao A, and Kan YW. Isolation of NF-E2-related factor 2 (Nrf2), a NF-E2-like basic leucine zipper transcriptional activator that binds to the tandem NF-E2/AP1 repeat of the beta-globin locus control region. *Proc Natl Acad Sci U S A* 91: 9926–9930, 1994.
136. Moncada S, Palmer RM, and Higgs EA. Biosynthesis of nitric oxide from L-arginine: a pathway for the regulation of cell function and communication. *Biochem Pharmacol* 38: 1709–1715, 1989.
137. Moncada S and Bolanos JP. Nitric oxide, cell bioenergetics and neurodegeneration. *J Neurochem* 97: 1676–1689, 2006.
138. Monti B, Berteotti C, and Contestabile A. Subchronic rolipram delivery activates hippocampal CREB and arc, enhances retention and slows down extinction of conditioned fear. *Neuropsychopharmacology* 31: 278–286, 2006.
139. Motohashi H and Yamamoto M. Nrf2-Keap1 defines a physiologically important stress response mechanism. *Trends Mol Med* 10: 549–557, 2004.
140. Motterlini R, Foresti R, Bassi R, Calabrese V, Clark JE, and Green CJ. Endothelial heme oxygenase-1 induction by hypoxia: modulation by inducible nitric-oxide synthase and S-nitrosothiols. *J Biol Chem* 275: 13613–13620, 2000.
141. Motterlini R, Green CJ, and Foresti R. Regulation of heme oxygenase-1 by redox signals involving nitric oxide. *Antiox Redox Signal* 4: 615–624, 2002.
142. Motterlini R, Green CJ, and Foresti R. Regulation of heme oxygenase-1 by redox signals involving nitric oxide. *Antiox Redox Signal* 4: 615–624, 2002.
143. Mungrue IN and Bredt DS. nNOS at a glance: implications for brain and brawn. *Cell Sci* 117: 2627–2629, 2004.
144. Murad F. Regulation of cytosolic guanylate cyclase by nitric oxide: the NO-cyclic GMP signal transduction system. *Adv Pharmacol* 26: 19–33, 1994.
145. Murphy MP, Packer MA, Scarlett JL, and Martin SW. Peroxynitrite: a biologically significant oxidant. *Gen Pharmacol* 31: 179–186, 1998.
146. Nakamura T and Lipton SA. Emerging roles of S-nitrosylation in protein misfolding and neurodegenerative diseases. *Antioxid Redox Signal* 10: 87–101, 2008.
147. Naughton P, Hoque M, Green CJ, Foresti R, and Motterlini R. Interaction of heme with nitroxyl or nitric oxide amplifies heme oxygenase-1 induction: involvement of the transcription factor Nrf2. *Cell Mol Biol (Noisy-le-grand)* 48: 885–894, 2002.
148. Nguyen T, Sherratt PJ, and Pickett CB. Regulatory mechanisms controlling gene expression mediated by the antioxidant response element. *Annu Rev Pharmacol Toxicol* 43: 233–260, 2003.
149. Nioi P, McMahon M, Itoh K, Yamamoto M, and Hayes JD. Identification of a novel Nrf2-regulated antioxidant response element (ARE) in the mouse NAD(P)H:quinone oxidoreductase 1 gene: reassessment of the ARE consensus sequence. *Biochem J* 374: 337–348, 2003.
150. Nott A, Watson PM, Robinson JD, Crepaldi L, and Riccio A. S-Nitrosylation of histone deacetylase 2 induces chromatin remodelling in neurons. *Nature* 455: 411–415, 2008.
151. Ojika K, Tsugu Y, Mitake S, Otsuka Y, and Katada E. NMDA receptor activation enhances the release of a cholinergic differentiation peptide (HCNP) from hippocampal neurons in vitro. *Brain Res Dev Brain Res* 106: 173–180, 1998.
152. Onofri F, Messa M, Matafora V, Bonanno G, Corradi A, Bachi A, Valtorta F, and Benfenati F. Synapsin phosphorylation by SRC tyrosine kinase enhances SRC activity in synaptic vesicles. *J Biol Chem* 282: 15754–15767, 2007.
153. Opie WO, Joshi G, Head E, Milgram NW, Muggenburg BA, Klein JB, Pierce WM, Cotman CW, and Butterfield DA. Proteomic identification of brain proteins in the canine model of human aging following a long-term treatment with antioxidants and a program of behavioral enrichment: relevance to Alzheimer's disease. *Neurobiol Aging* 29: 51–70, 2008.

154. Otori T, Friedland JC, Sinson G, McIntosh TK, Raghupathi R, and Welsh FA. Traumatic brain injury elevates glycogen and induces tolerance to ischemia in rat brain. *J Neurotrauma* 21: 707–718, 2004.
155. Owen JB, Di Domenico F, Sultana R, Perluigi M, Cini C, Pierce WM, and Butterfield DA. Proteomics-determined differences in the concanavalin-A-fractionated proteome of hippocampus and inferior parietal lobule in subjects with Alzheimer's disease and mild cognitive impairment: implications for progression of AD. *J Proteome Res* 8: 471–482, 2009.
156. Packer MA, Stasiv Y, Benraiss A, Chmielnicki E, Grinberg H, Westphal H, Goldman SA, and Enikolopov G. Nitric oxide negatively regulates mammalian adult neurogenesis. *Proc Natl Acad Sci U S A* 100: 9566–9571, 2003.
157. Pacher P, Schulz R, Liaudet L, and Szabo C. Nitrosative stress and pharmacological modulation of heart failure. *Trends Pharmacol Sci* 26: 302–310, 2005.
158. Pacher P, Beckman JS, and Liaudet L. Nitric oxide and peroxynitrite in health and disease. *Physiol Rev* 87: 315–424, 2007.
159. Paget MS and Buttner MJ. Thiol-based regulatory switches. *Annu Rev Genet* 37: 91–121, 2003.
160. Pantano C, Reynaert NL, van der Vliet A, and Janssen-Heininger YM. Redox-sensitive kinases of the nuclear factor-kappaB signaling pathway. *Antioxid Redox Signal* 8: 1791–1806, 2006.
161. Park S, Geddes TJ, Javitch JA, and Kuhn DM. Dopamine prevents nitration of tyrosine hydroxylase by peroxynitrite and nitrogen dioxide: is nitrotyrosine formation an early step in dopamine neuronal damage? *J Biol Chem* 278: 28736–28742, 2003.
162. Park SW, Huq MD, Hu X, and Wei LN. Tyrosine nitration on p65: a novel mechanism to rapidly inactivate nuclear factor-kappaB. *Mol Cell Proteomics* 4: 300–309, 2005.
163. Peraz-Mato I, Castro C, Ruiz FA, Corrales FJ, and Mato JM. Methionine adenosyltransferase S-nitrosylation is regulated by the basic and acidic amino acids surrounding the target thiol. *J Biol Chem* 274: 7075–7079, 1999.
164. Perkins ND. Integrating cell-signalling pathways with NF-kappaB and IKK function. *Nat Rev Mol Cell Biol* 8: 49–62, 2007.
165. Poderoso JJ, Carreras MC, Schopfer F, Lisdero C, Riobo N, Giulivi C, Boveris A, Boveris AA, and Cadenas E. The reaction of nitric oxide with ubiquinol: kinetic properties and biological significance. *Free Radic Biol Med* 26: 925–935, 1999.
166. Poon HF, Calabrese V, Calvani M, and Butterfield DA. Proteomics analyses of specific protein oxidation and protein expression in aged rat brain and its modulation by L-acetylcarnitine: insights into the mechanisms of action of this proposed therapeutic agent for CNS disorders associated with oxidative stress. *Antioxid Redox Signal* 8: 381–394, 2006.
167. Przedborski S and Ischiropoulos H. Reactive oxygen and nitrogen species: weapons of neuronal destruction in models of Parkinson's disease. *Antioxid Redox Signal* 7: 685–693, 2005.
168. Ramsey MM and Whiteley M. Polymicrobial interactions stimulate resistance to host innate immunity through metabolite perception. *Proc Natl Acad Sci U S A* 106: 1578–1583, 2009.
169. Reed TT, Owen J, Pierce WM, Sebastian A, Sullivan PG, and Butterfield DA. Proteomic identification of nitrated brain proteins in traumatic brain-injured rats treated post-injury with gamma-glutamylcysteine ethyl ester: insights into the role of elevation of glutathione as a potential therapeutic strategy for traumatic brain injury. *J Neurosci Res* 87: 408–417, 2009.
170. Reiter T. NO chemistry: a diversity of targets in the cell. *Redox Report* 11: 194–206, 2006.
171. Reuter S, Eifes S, Dicato M, Aggarwal BB, and Diederich M. Modulation of anti-apoptotic and survival pathways by curcumin as a strategy to induce apoptosis in cancer cells. *Biochem Pharmacol* 76: 1340–1351, 2008.
172. Rhee SG, Chae HZ, and Kim K. Peroxiredoxins: a historical overview and speculative preview of novel mechanisms and emerging concepts in cell signaling. *Free Radic Biol Med* 38: 1543–1552, 2005.
173. Riccio A, Alvania RS, Lonze BE, Ramanan N, Kim T, Huang Y, Dawson TM, Snyder SH, and Ginty DD. A nitric oxide signaling pathway controls CREB-mediated gene expression in neurons. *Mol Cell* 21: 283–294, 2006.
174. Rivier C. Role of gaseous neurotransmitters in the hypothalamic-pituitary-adrenal axis. *Ann N Y Acad Sci* 933: 254–264, 2001.
175. Rosenberg PA, Li Y, Ali S, Altiock N, Back SA, and Volpe JJ. Intracellular redox state determines whether nitric oxide is toxic or protective to rat oligodendrocytes in culture. *J Neurochem* 73: 476–484, 1999.
176. Rushmore TH and Pickett CB. Transcriptional regulation of the rat glutathione S-transferase Ya subunit gene: characterization of a xenobiotic-responsive element controlling inducible expression by phenolic antioxidants. *J Biol Chem* 265: 14648–14653, 1990.
177. Rushmore TH, Morton MR, and Pickett CB. The antioxidant responsive element: activation by oxidative stress and identification of the DNA consensus sequence required for functional activity. *J Biol Chem* 266: 11632–11639, 1991.
178. Saha RN and Pahan K. Regulation of inducible nitric oxide synthase gene in glial cells. *Antioxid Redox Signal* 8: 929–947, 2006.
179. Sakiyama T, Musch MW, Ropeleski MJ, Tsubouchi H, and Chang EB. Glutamine increases autophagy under basal and stressed conditions in intestinal epithelial cells. *Gastroenterology* 136: 924–932, 2009.
180. Satoh T and Lipton SA. Redox regulation of neuronal survival mediated by electrophilic compounds. *Trends Neurosci* 1: 37–45, 2007.
181. Sawa T, Zaki MH, Okamoto T, Akuta T, Tokutomi Y, Kim-Mitsuyama S, Ihara H, Kobayashi A, Yamamoto M, Fujii S, Arimoto H, and Akaike T. Protein S-guanylation by the biological signal 8-nitroguanosine 3', 5'-cyclic monophosphate. *Nat Chem Biol* 3: 727–735, 2007.
182. Scapagnini G, Foresti R, Calabrese V, Giuffrida Stella AM, Green CJ, and Motterlini R. Caffeic acid phenethyl ester and curcumin: a novel class of heme oxygenase-1 inducers. *Mol Pharmacol* 61: 554–561, 2002.
183. Scapagnini G, Giuffrida Stella AM, Abraham NG, Alkon D, and Calabrese V. Differential expression of heme oxygenase-1 in rat brain by endotoxin (LPS). In: *Heme oxygenase in biology and medicine*. Abraham NG, Alkon D, and Calabrese V. eds. New York: Kluwer Academic Publisher, 2002, pp. 121–134.
184. Schwarzbach E, Bonislawski DP, Xiong G, and Cohen AS. Mechanisms underlying the inability to induce area CA1 LTP in the mouse after traumatic brain injury. *Hippocampus* 16: 541–550, 2006.
185. Shalova IN, Cechalova K, Rehakova Z, Dimitrova P, Og-nibene E, Caprioli A, Schmalhausen EV, Muronetz VI, and

- Saso L. Decrease of dehydrogenase activity of cerebral glyceraldehyde-3-phosphate dehydrogenase in different animal models of Alzheimer's disease. *Biochim Biophys Acta* 1770: 826–832, 2007.
186. Shelton MD and Miesal JJ. Regulation by reversible S-glutathionylation: molecular targets implicated in inflammatory diseases. *Mol Cell* 25: 332–346, 2008.
 187. Sliskovic I, Raturi A, and Mutus B. Characterization of the S-denitrosation activity of protein disulfide isomerase. *J Biol Chem* 280: 8733–8741, 2005.
 188. Smith GS, Kramer E, Ma Y, Hermann CR, Dhawan V, Chaly T, and Eidelberg D. Cholinergic modulation of the cerebral metabolic response to citalopram in Alzheimer's disease. *Brain* 132: 392–401, 2009.
 189. Son H, Hawkins RD, Martin K, Kiebler M, Huang PL, Fishman MC, and Kandel ER. Long-term potentiation is reduced in mice that are doubly mutant in endothelial and neuronal nitric oxide synthase. *Cell* 87: 1015–1023, 1996.
 190. Spiro S. Regulators of bacterial responses to nitric oxide. *FEMS Microbiol Rev* 31: 193–211, 2007.
 191. Squadrito GL and Pryor WA. Oxidative chemistry of nitric oxide: the roles of superoxide, peroxynitrite, and carbon dioxide. *Free Radic Biol Med* 55: 392–403, 1998.
 192. Stadtman ER, Moskovitz J, and Levine RL. Oxidation of methionine residues of proteins: biological consequences. *Antioxid Redox Signal* 5: 577–582, 2003.
 193. Stamler JS. Redox signaling: nitrosylation and related target interactions of nitric oxide. *Cell* 78: 931–936, 1994.
 194. Stanyer L, Jorgensen W, Hori O, Clark JB, and Heales SJ. Inactivation of brain mitochondrial Lon protease by peroxynitrite precedes electron transport chain dysfunction. *Neurochem Int* 3: 95–101, 2008.
 195. Sultana R, Poon HF, Cai J, Pierce WM, Merchant M, Klein JB, Markesbery WR, and Butterfield DA. Identification of nitrated proteins in Alzheimer's disease brain using a redox proteomics approach. *Neurobiol Dis* 22: 76–87, 2006.
 196. Sultana R, Reed T, Perluigi M, Coccia R, Pierce WM, and Butterfield DA. Proteomic identification of nitrated brain proteins in amnesic mild cognitive impairment: a regional study. *J Cell Mol Med* 11: 839–851, 2007.
 197. Sultana R, Boyd-Kimball D, Cai J, Pierce WM, Klein JB, Merchant M, and Butterfield DA. Proteomics analysis of the Alzheimer's disease hippocampal proteome. *J Alzheimers Dis* 11: 153–164, 2007.
 198. Sun XF and Zhang H. NFKB and NFKBI polymorphisms in relation to susceptibility of tumour and other diseases. *Histol Histopathol* 22: 1387–1398, 2007.
 199. Talalay P, Fahey JW, Holtzclaw WD, Prestera T, and Zhang Y. Chemoprotection against cancer by phase 2 enzyme induction. *Toxicol Lett* 83: 173–179, 1995.
 200. Talalay P. Chemoprotection against cancer by induction of phase 2 enzymes. *Biofactors* 12: 5–11, 2000.
 201. Tannenbaum SR and White FM. Regulation and specificity of S-nitrosylation and denitrosylation. *ACS Chem Biol* 1: 615–618, 2006.
 202. Trabace L, Cassano T, Tucci P, Steardo L, Kendrick KM, and Cuomo V. The effects of nitric oxide on striatal serotonergic transmission involve multiple targets: an in vivo microdialysis study in the awake rat. *Brain Res* 2008: 293–298, 2004.
 203. Tsang AH, Lee YI, Ko HS, Savitt JM, Pletnikova O, Troncoso JC, Dawson VL, Dawson TM, and Chung KK. S-Nitrosylation of XIAP compromises neuronal survival in Parkinson's disease. *Proc Natl Acad Sci U S A* 106: 4900–4905, 2009.
 204. Uehara T, Nakamura T, Yao D, Shi ZQ, Gu Z, Ma Y, Masliah E, Nomura Y, and Lipton SA. S-Nitrosylated protein-disulphide isomerase links protein misfolding to neurodegeneration. *Nature* 441: 513–517, 2006.
 205. Vesela A and Wilhelm J. The role of carbon dioxide in free radical reactions of the organism. *Physiol Res* 51: 335–339, 2002.
 206. Voelker R. Parkinson disease guidelines aid diagnosis, management. *JAMA* 295: 2126–2128, 2006.
 207. Wakabayashi N, Dinkova-Kostova AT, Holtzclaw WD, Kang MI, Kobayashi A, Yamamoto M, Kensler TW, and Talalay P. Protection against electrophile and oxidant stress by induction of the phase 2 response: fate of cysteines of the Keap1 sensor modified by inducers. *Proc Natl Acad Sci U S A* 101: 2040–2045, 2004.
 208. Wang HD, Lu DX, and Qi RB. Therapeutic strategies targeting the LPS signaling and cytokines. *Pathophysiology* 2009 Mar 23. [Epub ahead of print].
 209. Williams JL, Kashfi K, Ouyang N, del Soldato P, Kopelovich L, and Rigas B. NO-donating aspirin inhibits intestinal carcinogenesis in Min (APC(Min/+)) mice. *Biochem Biophys Res Commun* 313: 784–748, 2004.
 210. Xu L, Eu JP, Meissner G, and Stamler JS. Activation of the cardiac calcium release channel (ryanodine receptor) by poly-S-nitrosylation. *Science* 279: 234–237, 1998.
 211. Xu S, Ying J, Jiang B, Guo W, Adachi T, Sharov V, Lazar H, Menzoian J, Knyushko TV, Bigelow D, Schoneich C, and Cohen RA. Detection of sequence-specific tyrosine nitration of manganese SOD and SERCA in cardiovascular disease and aging. *Am J Physiol Heart Circ Physiol* 290: H2220–H2227, 2006.
 212. Yao D, Gu Z, Nakamura T, Shi ZQ, Ma Y, Gaston B, Palmer LA, Rockenstein EM, Zhang Z, Masliah E, Uehara T, and Lipton SA. Nitrosative stress linked to sporadic Parkinson's disease: S-nitrosylation of parkin regulates its E3 ubiquitin ligase activity. *Proc Natl Acad Sci U S A* 101: 10810–10814, 2004.
 213. Yokoyama H, Takagi S, Watanabe Y, Kato H, and Araki T. Role of reactive nitrogen and reactive oxygen species against MPTP neurotoxicity in mice. *J Neural Transm* 115: 831–842, 2008.
 214. Yokoyama H, Yano R, Aoki E, Kato H, and Araki T. Comparative pharmacological study of free radical scavenger, nitric oxide synthase inhibitor, nitric oxide synthase activator and cyclooxygenase inhibitor against MPTP neurotoxicity in mice. *Metab Brain Dis* 23: 335–349, 2008.
 215. Zhang D, Shen J, Wang C, Zhang X, and Chen J. GSH-dependent iNOS and HO-1 mediated apoptosis of human Jurkat cells induced by nickel(II). *Environ Toxicol* 2008 epub before print.
 216. Zhang DD and Hannink M. Distinct cysteine residues in Keap1 are required for Keap1-dependent ubiquitination of Nrf2 and for stabilization of Nrf2 by chemopreventive agents and oxidative stress. *Mol Cell Biol* 23: 8137–8151, 2003.
 217. Zhang DD, Lo SC, Cross JV, Templeton DJ, and Hannink M. Keap1 is a redox-regulated substrate adaptor protein for a Cul3-dependent ubiquitin ligase complex. *Mol Cell Biol* 24: 10941–10953, 2004.
 218. Zhang H, Squadrito GL, Uppu RM, Lemerrier JN, Cueto R, and Pryor WA. Inhibition of peroxynitrite-mediated oxidation of glutathione by carbon dioxide. *Arch Biochem Biophys* 339: 183–189, 1997.

219. Zheng M, Aslund F, and Storz G. Activation of the OxyR transcription factor by reversible disulfide bond formation. *Science* 279: 1718–1721, 1998.
220. Zheng M and Storz G. Redox sensing by prokaryotic transcription factors. *Biochem Pharmacol* 59: 1–6, 2000.
221. Zhou P, Qian L, and Iadecola C. Nitric oxide inhibits caspase activation and apoptotic morphology but does not rescue neuronal death. *J Cereb Blood Flow Metab* 25: 348–357, 2005.

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Abbreviations Used

AD = Alzheimer's disease
Akt = protein kinase B
ALS = amyotrophic lateral sclerosis
AP-1 = activator protein-1
ARE/EpRE = antioxidant/electrophile response element
BDNF = brain-derived neurotrophic factor
BH4 = tetrahydrobiopterin
cGMP = cyclic guanosine monophosphate
CNS = central nervous system
CREB = cAMP-response element-binding protein
DRP2 = dihydropyrimidase like-2
EC-SOD = extracellular SOD
EDRF = endothelium-derived relaxation factor
ER = endoplasmic reticulum
GABA = γ -aminobutyric acid
GAPDH = glyceraldehyde 3-phosphate dehydrogenase
GDP1 = guanosine diphosphate dissociation inhibitor 1
GRP = glucose-regulated protein
GSH = reduced glutathione
GSNO = S-nitrosoglutathione
GSSG = oxidized glutathione
GST = glutathione S-transferase
HCNP = hippocampal cholinergic neurostimulating peptide
HD = Huntington's disease
HDAC2 = histone deacetylase 2
Ho-1 = heme oxygenase 1

Hsp = heat-shock protein
IFN- γ = interferon- γ
Keap1 = Kelch-like ECH-associated protein 1
LTP = long-term potentiation
LTD = long-term depression
MCI = mild cognitive impairment
MMP9 = matrix metalloproteinase 9
MPTP = 1-methyl-4-phenyl-1,2,3,6-tetrahydropyridine
MRP3 = multidrug-resistance protein 3
MsrA = methionine sulfoxide reductase
mtNOS = mitochondrial NOS
NAD(P)H = reduced nicotinamide adenine dinucleotide phosphate
NF- κ B = nuclear factor- κ B
7-NI = 7-nitroindazole
8-nitro-cGMP = 8-nitroguanosine 3',5'-cyclic monophosphate
NMDA = N-methyl-D-aspartate
NO = nitric oxide
NOS = nitric oxide synthase
NQO1 = NAD(P)H:quinone oxidoreductase 1
O₂ = molecular oxygen
ODQ = 1H-[1,2,4]oxadiazolo[4,3-a]quinoxalin-1-one
ONOO- = peroxynitrite
PD = Parkinson's disease
PDI = protein-disulfide isomerase
PDK1 = phosphatidylinositol-dependent kinase 1
PEBP = phosphatidylethanolamine-binding protein
PGM = phosphoglycerate mutase
PI3K = phosphatidylinositol 3-kinase
PIP3 = phosphatidylinositol-3,4,5-trisphosphate
PKG = protein kinase G
PRX2 = peroxiredoxin 2
PSSG = protein-mixed disulfides/glutathionylated cysteine
PUFAs = polyunsaturated fatty acids
RKIP = raf-kinase inhibitor protein
RNS = reactive nitrogen species
ROS = reactive oxygen species
8-RS-cGMP = 8-thioalkoxy-cGMP
RSNO = S-nitrosothiol
SERCA = sarcoplasmic reticulum Ca²⁺ ATPase
sGC = soluble guanylate cyclase
SNO = S-nitrosylation, S-nitrosothiol, S-nitrosylated protein
SOD = superoxide dismutase
SOH = sulfenic acid
SO₂H = sulfinic acid
S-Ox = oxidized cysteine
Srx = sulfiredoxin
S-S = disulfide
TBI = traumatic brain injury
YC-1 = 3-(5-hydroxymethyl-2-furyl)-1-benzyl-indazole

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1. Derek A. Drechsel, Alvaro G. Estévez, Luis Barbeito, Joseph S. Beckman. 2012. Nitric Oxide-Mediated Oxidative Damage and the Progressive Demise of Motor Neurons in ALS. *Neurotoxicity Research* **22**:4, 251-264. [[CrossRef](#)]
2. Alexander A. Mongin , Preeti Dohare , David Jour'dheuil . 2012. Selective Vulnerability of Synaptic Signaling and Metabolism to Nitrosative Stress. *Antioxidants & Redox Signaling* **17**:7, 992-1012. [[Abstract](#)] [[Full Text HTML](#)] [[Full Text PDF](#)] [[Full Text PDF with Links](#)]
3. Massimo Bertinaria, Barbara Rolando, Marta Giorgis, Gabriele Montanaro, Elisabetta Marini, Massimo Collino, Elisa Benetti, Pier Giuseppe Daniele, Roberta Fruttero, Alberto Gasco. 2012. Carnosine analogues containing NO-donor substructures: Synthesis, physico-chemical characterization and preliminary pharmacological profile. *European Journal of Medicinal Chemistry* **54**, 103-112. [[CrossRef](#)]
4. Jiangang Long, Vadim Aksenov, C. David Rollo, Jiankang Liu. 2012. A complex dietary supplement modulates nitrative stress in normal mice and in a new mouse model of nitrative stress and cognitive aging. *Mechanisms of Ageing and Development* **133**:8, 523-529. [[CrossRef](#)]
5. Milos R. Filipovic, Jan Lj. Miljkovic, Thomas Nauser, Maksim Royzen, Katharina Klos, Tatyana Shubina, Willem H. Koppenol, Stephen J. Lippard, Ivana Ivanovi#-Burmazovi#. 2012. Chemical Characterization of the Smallest S -Nitrosothiol, HSNO; Cellular Cross-talk of H₂S and S -Nitrosothiols. *Journal of the American Chemical Society* **134**:29, 12016-12027. [[CrossRef](#)]
6. Anil K. Mantha, Monisha Dhiman, Giulio Taglialatela, Regino J. Perez-Polo, Sankar Mitra. 2012. Proteomic study of amyloid beta (25-35) peptide exposure to neuronal cells: Impact on APE1/Ref-1's protein-protein interaction. *Journal of Neuroscience Research* **90**:6, 1230-1239. [[CrossRef](#)]
7. Neelam Shahani, Akira Sawa. 2012. Protein S-nitrosylation: Role for nitric oxide signaling in neuronal death. *Biochimica et Biophysica Acta (BBA) - General Subjects* **1820**:6, 736-742. [[CrossRef](#)]
8. Jun-Ha Hwang, Sung Ah Moon, Cham Han Lee, Mi Ran Byun, A Rum Kim, Mi Kyung Sung, Hyun-Jin Park, Eun Sook Hwang, Sang Hyun Sung, Jeong-Ho Hong. 2012. Idesolide inhibits the adipogenic differentiation of mesenchymal cells through the suppression of nitric oxide production. *European Journal of Pharmacology* **685**:1-3, 218-223. [[CrossRef](#)]
9. Elizabeth A. Sabens Liedhegner , Xing-Huang Gao , John J. Mieyal . 2012. Mechanisms of Altered Redox Regulation in Neurodegenerative Diseases—Focus on S-Glutathionylation. *Antioxidants & Redox Signaling* **16**:6, 543-566. [[Abstract](#)] [[Full Text HTML](#)] [[Full Text PDF](#)] [[Full Text PDF with Links](#)]
10. Volodymyr I. Lushchak. 2012. Glutathione Homeostasis and Functions: Potential Targets for Medical Interventions. *Journal of Amino Acids* **2012**, 1-26. [[CrossRef](#)]
11. M. Alba Sorolla, María José Rodríguez-Colman, Núria Vall-Illaura, Jordi Tamarit, Joaquim Ros, Elisa Cabiscol. 2012. Protein oxidation in Huntington disease. *BioFactors* n/a-n/a. [[CrossRef](#)]
12. El#bieta Miller, Anna Walczak, Joanna Saluk, Micha# B. Ponczek, Ireneusz Majsterek. 2012. Oxidative modification of patient's plasma proteins and its role in pathogenesis of multiple sclerosis. *Clinical Biochemistry* **45**:1-2, 26-30. [[CrossRef](#)]
13. Bruno P. Carreira, Caetana M. Carvalho, Inês M. Araújo. 2012. Regulation of Injury-Induced Neurogenesis by Nitric Oxide. *Stem Cells International* **2012**, 1-15. [[CrossRef](#)]
14. Zsolt Radak, Hisashi Naito, Albert W. Taylor, Sataro Goto. 2011. Nitric oxide: Is it the cause of muscle soreness?. *Nitric Oxide* . [[CrossRef](#)]
15. Changgong Wu , Andrew M. Parrott , Cexiong Fu , Tong Liu , Stefano M. Marino , Vadim N. Gladyshev , Mohit R. Jain , Ahmet T. Baykal , Qing Li , Shinichi Oka , Junichi Sadoshima , Annie Beuve , William J. Simmons , Hong Li . 2011. Thioredoxin 1-Mediated Post-Translational Modifications: Reduction, Transnitrosylation, Denitrosylation, and Related Proteomics Methodologies. *Antioxidants & Redox Signaling* **15**:9, 2565-2604. [[Abstract](#)] [[Full Text HTML](#)] [[Full Text PDF](#)] [[Full Text PDF with Links](#)] [[Supplemental material](#)]
16. Vittorio Calabrese, Carolin Cornelius, Albena T. Dinkova-Kostova, Ivo Iavicoli, Rosanna Di Paola, Aleardo Koverech, Salvatore Cuzzocrea, Enrico Rizzarelli, Edward J. Calabrese. 2011. Cellular stress responses, hormetic phytochemicals and vitagenes in aging and longevity. *Biochimica et Biophysica Acta (BBA) - Molecular Basis of Disease* . [[CrossRef](#)]
17. G. Pennisi, C. Cornelius, M.M. Cavallaro, A. Trovato Salinaro, M.T. Cambria, M. Pennisi, R. Bella, P. Milone, B. Ventimiglia, M.R. Migliore, L. Di Renzo, A. De Lorenzo, V. Calabrese. 2011. Redox regulation of cellular stress response in multiple sclerosis. *Biochemical Pharmacology* **82**:10, 1490-1499. [[CrossRef](#)]
18. D R Gough, T G Cotter. 2011. Hydrogen peroxide: a Jekyll and Hyde signalling molecule. *Cell Death and Disease* **2**:10, e213. [[CrossRef](#)]

19. Vittorio Calabrese, Carolin Cornelius, Salvatore Cuzzocrea, Ivo Iavicoli, Enrico Rizzarelli, Edward J. Calabrese. 2011. Hormesis, cellular stress response and vitagenes as critical determinants in aging and longevity. *Molecular Aspects of Medicine* . [[CrossRef](#)]
20. Yukio Ago, Toshiyuki Kawasaki, Tetsuaki Nashida, Yuki Ota, Yana Cong, Mari Kitamoto, Teisuke Takahashi, Kazuhiro Takuma, Toshio Matsuda. 2011. SEA0400, a specific Na⁺/Ca²⁺ exchange inhibitor, prevents dopaminergic neurotoxicity in an MPTP mouse model of Parkinson's disease. *Neuropharmacology* . [[CrossRef](#)]
21. Hao Yu, Thomas J. Payne, Dillip K. Mohanty. 2011. Effects of Slow, Sustained, and Rate-Tunable Nitric Oxide Donors on Human Aortic Smooth Muscle Cells Proliferation. *Chemical Biology & Drug Design* no-no. [[CrossRef](#)]
22. Rodrigo Sandoval, Andrés González, Ariel Caviedes, Floria Pancetti, Karl-Heinz Smalla, Thilo Kaehne, Luis Michea, Eckart D. Gundelfinger, Ursula Wyneken. 2011. Homeostatic NMDA receptor down-regulation via brain derived neurotrophic factor and nitric oxide-dependent signalling in cortical but not in hippocampal neurons. *Journal of Neurochemistry* no-no. [[CrossRef](#)]
23. S. Li, A.C. Vana, R. Ribeiro, Y. Zhang. 2011. Distinct role of nitric oxide and peroxynitrite in mediating oligodendrocyte toxicity in culture and in experimental autoimmune encephalomyelitis. *Neuroscience* **184**, 107-119. [[CrossRef](#)]
24. Kh. L. Gainutdinov, S. A. Gavrilova, V. S. Iyudin, A. V. Golubeva, M. P. Davydova, G. G. Jafarova, V. V. Andrianov, V. B. Koshelev. 2011. EPR Study of the Intensity of the Nitric Oxide Production in Rat Brain After Ischemic Stroke. *Applied Magnetic Resonance* **40**:3, 267-278. [[CrossRef](#)]
25. Neelam Shahani , Akira Sawa . 2011. Nitric Oxide Signaling and Nitrosative Stress in Neurons: Role for S-Nitrosylation. *Antioxidants & Redox Signaling* **14**:8, 1493-1504. [[Abstract](#)] [[Full Text HTML](#)] [[Full Text PDF](#)] [[Full Text PDF with Links](#)]
26. Frank B. Jensen, Marie N. Hansen. 2011. Differential uptake and metabolism of nitrite in normoxic and hypoxic goldfish. *Aquatic Toxicology* **101**:2, 318-325. [[CrossRef](#)]
27. Young-Myeong Kim , Hyun-Ock Pae , Jeong Euy Park , Yong Chul Lee , Je Moon Woo , Nam-Ho Kim , Yoon Kyung Choi , Bok-Soo Lee , So Ri Kim , Hun-Taeg Chung . 2011. Heme Oxygenase in the Regulation of Vascular Biology: From Molecular Mechanisms to Therapeutic Opportunities. *Antioxidants & Redox Signaling* **14**:1, 137-167. [[Abstract](#)] [[Full Text HTML](#)] [[Full Text PDF](#)] [[Full Text PDF with Links](#)]
28. Young-Don Kwak, Ruishan Wang, Jing Li, Yun-Wu Zhang, Huaxi Xu, Francesca-Fang Liao. 2011. Differential regulation of BACE1 expression by oxidative and nitrosative signals. *Molecular Neurodegeneration* **6**:1, 17. [[CrossRef](#)]
29. Vittorio Calabrese , Carolin Cornelius , Alben T. Dinkova-Kostova , Edward J. Calabrese , Mark P. Mattson . 2010. Cellular Stress Responses, The Hormesis Paradigm, and Vitagenes: Novel Targets for Therapeutic Intervention in Neurodegenerative Disorders. *Antioxidants & Redox Signaling* **13**:11, 1763-1811. [[Abstract](#)] [[Full Text HTML](#)] [[Full Text PDF](#)] [[Full Text PDF with Links](#)]
30. Vittorio Calabrese, Carolin Cornelius, Anna Maria Giuffrida Stella, Edward J. Calabrese. 2010. Cellular Stress Responses, Mitostress and Carnitine Insufficiencies as Critical Determinants in Aging and Neurodegenerative Disorders: Role of Hormesis and Vitagenes. *Neurochemical Research* **35**:12, 1880-1915. [[CrossRef](#)]
31. Vittorio Calabrese, C. Cornelius, L. Maiolino, M. Luca, R. Chiamonte, M. A. Toscano, A. Serra. 2010. Oxidative Stress, Redox Homeostasis and Cellular Stress Response in Ménière's Disease: Role of Vitagenes. *Neurochemical Research* **35**:12, 2208-2217. [[CrossRef](#)]
32. M. Perluigi, F. Di Domenico, A. Giorgi, M.E. Schinina, R. Coccia, C. Cini, F. Bellia, M.T. Cambria, C. Cornelius, D.A. Butterfield, V. Calabrese. 2010. Redox proteomics in aging rat brain: Involvement of mitochondrial reduced glutathione status and mitochondrial protein oxidation in the aging process. *Journal of Neuroscience Research* **88**:16, 3498-3507. [[CrossRef](#)]
33. Giuseppe Caravaglios, Giuseppe Castro, Erminio Costanzo, Giulia Maria, Danielle Mancuso, Emma Gabriella Muscoso. 2010. Theta power responses in mild Alzheimer's disease during an auditory oddball paradigm: lack of theta enhancement during stimulus processing. *Journal of Neural Transmission* **117**:10, 1195-1208. [[CrossRef](#)]
34. Rainer Metternich, Giorgio Tarzia. 2010. "Hot Spots" in Medicinal Chemistry. *ChemMedChem* **5**:8, 1159-1162. [[CrossRef](#)]
35. Mayumi Kajimura, Ryo Fukuda, Ryon M. Bateman, Takehiro Yamamoto, Makoto Suematsu . 2010. Interactions of Multiple Gas-Transducing Systems: Hallmarks and Uncertainties of CO, NO, and H₂S Gas Biology. *Antioxidants & Redox Signaling* **13**:2, 157-192. [[Abstract](#)] [[Full Text HTML](#)] [[Full Text PDF](#)] [[Full Text PDF with Links](#)]
36. Annadora J. Bruce-Keller , Sunita Gupta , Taryn E. Parrino , Alecia G. Knight , Philip J. Ebenezer , Adam M. Weidner , Harry LeVine III , Jeffrey N. Keller , William R. Markesbery . 2010. NOX Activity Is Increased in Mild Cognitive Impairment. *Antioxidants & Redox Signaling* **12**:12, 1371-1382. [[Abstract](#)] [[Full Text HTML](#)] [[Full Text PDF](#)] [[Full Text PDF with Links](#)]
37. Soo-Jin Oh, Jee-In Heo, Yoon-Jung Kho, Jeong-Hyeon Kim, Hong-Joon Kang, Seong-Hoon Park, Hyun-Seok Kim, Jong-Yeon Shin, Min-Ju Kim, Sung Chan Kim, Jae-Bong Park, Jaebong Kim, Jae-Yong Lee. 2010. Nitric Oxide Is an Essential Mediator for Neuronal Differentiation of Rat Primary Cortical Neuron Cells. *Experimental Neurobiology* **19**:2, 83. [[CrossRef](#)]

38. Paola Chiarugi . 2009. Survival or Death: The Redox Paradox. *Antioxidants & Redox Signaling* **11**:11, 2651-2654. [[Citation](#)]
[[Full Text HTML](#)] [[Full Text PDF](#)] [[Full Text PDF with Links](#)]